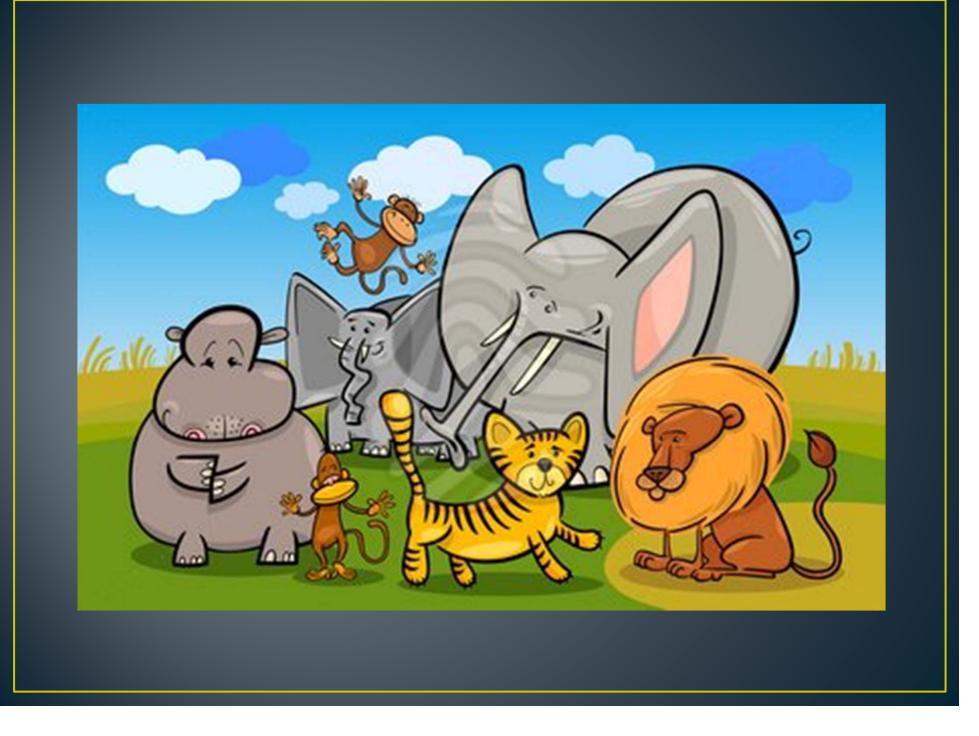
# Validation of Diagnostic Tests for Wildlife

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# Brucella melitensis is absent



Serology (i-ELISA) for Brucella melitensis: 10/25 animals test positive



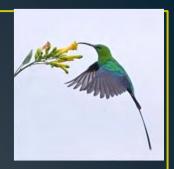


#### Manufacturer's specifications:

- Indirect ELISA • Method:
- Species: Ruminants
- Specimens: Bovine, ovine and caprine serum

#### Test not validated for the target species!

# What is test validation?



- Process that determines the fitness of an assay for an intended purpose
- The assay is subjected to a validation pathway in which the assay's analytical and diagnostic performance is determined.

#### Note:

Test validation depends on compliance with Quality management in veterinary testing laboratories (e.g. ISO17025)



Confidence in test results obtained

Ensure quality of the test results



### Diagnostic tests Fitness for (intended) purpose

The capacity of a positive or negative test result to predict accurately the infection or exposure status of the animal or population of animals is the ultimate consideration of assay validation.











Diagnostic testing of wildlife becoming 1 important **because of** diseases which can have an impact on

- wildlife populations and biodiversity
- Health of humans
- Health of domestic animals

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Approach adopted for test validation in domestic animals presents challenges for use in wildlife



Develop approach applicable to wildlife

### Intended purposes (wildlife)

- 1) Screening wildlife populations for the presence of infectious agents, for example:
  - a) for surveillance (e.g. early detection, evaluation of trends in prevalence or incidence)
  - b) to estimate prevalence of infection or exposure
- 2) Screening or testing vectors or environmental samples for the presence of infectious agents
- 3) Confirming a diagnosis of suspect or clinical cases (includes confirmation of positive results from a screening test)
- 4) Certifying freedom from infection or presence of the agent in individual animals or products, for
- a) movement or translocation
- b) human consumption
- 5) Monitoring of the geographical distribution and prevalence changes due to management interventions (including determining immune status of individual animals or populations)
- 6) Studying agent, host and environment factors associated with disease occurrence

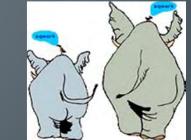
### The road to test validation

#### Characteristics used in validation

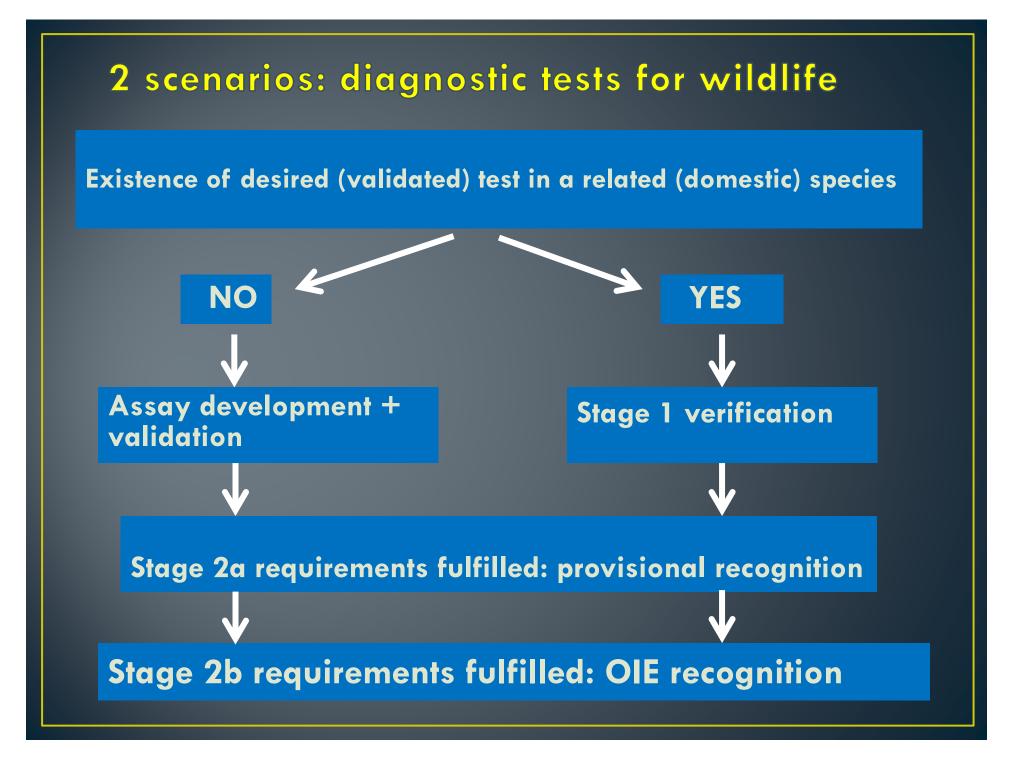
- Definition of the intended purpose(s)
- Optimization
- Standardisation
- Analytical sensitivity
- Analytical specificity
- Diagnostic sensitivity
- Diagnostic specificity
- Thresholds (cut-offs)
- Reproducibility







Assay development pathway	Definition of intended purpose	Study design + protocol	Reagents + controls Optimisation
Assay validation pathway	Analytical sensitivity	Stage 1: Analytical	Repeatability and preliminary reproducibility
	Analytical specificity	characteristics	Candidate test compared with reference test
	Diagnostic sensitivity		Samples from reference animals
	Diagnostic specificity	Stage 2: Diagnostic characteristics	Samples from experimental animals
	Cut-off determination		
	Select collaborating labs	Stage 3: Reproducibility —	Assay designated as • "validated for the original intended purpose"
	Deploy to other labs	← Stage 4: Implementation	Reference standards selected
		<	Internat.recognition by OIE
Validation Status Retention	Assay modifications and re-validation	Monitoring and maintenance of validation criteria	Daily in-house QC Proficiency testing





Test development pathway	Definition of intended purpose	Study design + protocol Optimisation etc.
Test validation pathway	Analytical sensitivity	Repeatability and preliminary Stage 1: Analytical reproducibility
	Analytical specificity	characteristics Candidate test compared with reference test
Stage 1 + Stage 2a fulfilled: Provisional	Diagnostic sensitivity	
recognition	Diagnostic specificity	Stage 2a: Diagnostic characteristics -limited Characteristics -limited
	Cut-off determination	
LOCAL ACCEPTANCE for use in populations on national and	Diagnostic sensitivity	Stage 2b: Diagnostic
possibly regional level	Diagnostic specificity	characteristics -full reference samples
	Cut-off determination	
Internationalisation	Select collaborating labs	Stage 3: reproducibility Designated as "Validated for the original intended purpose"
	Deploy to other labs	Stage 4: implementation Internat.recognition by OIE

Validation pathway	Pathway 1: No validated test in	Pathway 2: Validated test in related	
Chapter 1.1.5.	related species*	species*	
Stage 1	Stage-1 verified in new target species	Stage-1 verified in new target species	
Analytical specificity	Yes	Yes	
Analytical sensitivity	Yes	Yes	
Repeatability	Yes	No	
Reproducibility (preliminary)	Yes	No	
Stage 2	Stage 2a (Provisional recognition)	Stage 2a (Provisional recognition)	
	Yes	Yes	
Diagnostic sensitivity	(minimum of <b>30</b> positive reference	(minimum of <b>10</b> positive reference	
	samples)	samples)	
Diagnostic specificity	Yes (minimum of <mark>30 negative</mark> reference	Yes (minimum of <b>10 negative</b> reference	
	samples)	samples)	
Cut-off determination	Yes (total of <b>60</b> samples)	Yes (total of <b>20</b> samples)	
Reference sample description	Yes	Yes	
	Stage 2b	Stage 2b	
Diagnostic sensitivity	Yes	Yes	
Diagnostic specificity	Yes	Yes	
Cut-off determination	Yes	Yes	
Reference sample description	Yes	Yes	
Stage 3	Stage 3	Stage 3	
Reproducibility	Yes	Yes	
Repeatability	Yes	Yes	
Stage 4	Stage 4	Stage 4	
Predictive values (populations)	Yes	Yes	

### Sample size for full validation: stage 2b - Domestic animals

Table 1. Theoretical number of samples from animals of known infection status required for establishing diagnostic sensitivity (DSe) and specificity (DSp) estimates with known confidence

<b>v</b> v						
	2% error allowed in estimate of DSe and DSp				5% error allowed in estimate of DSe and DSp	
Estimated		Confidence			Confidence	
DSe or DSp	90% 95% 99% 90% 95%		99%			
90%	610	864	1493	98	138	239
<b>92</b> %	466	707	1221	75	113	195
94%	382	542	935	61	87	150
<b>95</b> %	372	456	788	60	73	126
<b>96</b> %	260	369	637	42	59	102
<b>97</b> %	197	279	483	32	45	77
<b>98</b> %	133	188	325	21	30	52
<b>99</b> %	67	95	164	11	15	26

Percent error allowed in the estimate of DSe or DSp = 2% in the left panel and 5% in the right panel. For the number of samples required for 1%, 3%, and 4% error margin in the estimate of DSe and DSp, multiply the number of samples in the left panel of the table by a factor of 4.0, 0.44, and 0.25, respectively.

### Sample size for full validation: stage 2b - Wildlife

• Absolute numbers of samples may initially be lower

Calculated error margins will be wider

• Increase in uncertainty in test performance criteria

No. positive reference samples	No. positive	DSe (%)	Approximate error margin on estimate of DSe	95% exact binomial confidence interval for DSe (%)
140	126	90	± 0.05	83.8 - 94.4
100	90	90	± 0.06	82.4 – 95.1
60	54	90	± 0.08	79.5 – 96.2
30	27	90	± 0.10	73.5 – 97.9
10	9	90	± 0.18	55.5 – 99.7

### Sample size for full validation: stage 2b - Wildlife

#### Recommendations to obtain required certainty:

- Combine data from multiple laboratories
- Build validation database over time

#### **Recommendations cont'd:**

Using representative samples of the target condition is of greater importance than sample size.

### • Why?

 Achieves an unbiased (and practically useful) estimate of DSe and DSp that will stand up to scrutiny over time.

### How target conditions influence Dse

Johne's disease in cattle and sheep: < 5% show clinical signs

Respiratory infections: 50% clinically apparent

Anthrax: 98% are clinically apparent

# Common constraints





"This test is validated"



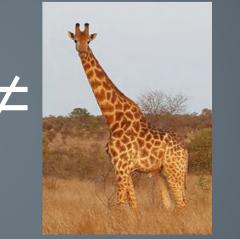
"This test is fit for purpose"

# Test constraints

### Indirect tests

#### Require species specific immunological reagents (antibodies)











# Test constraints A bit of good news...

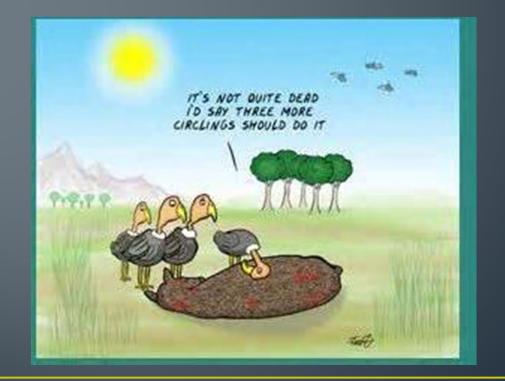
 Test methods for direct pathogen detection are generally not species-specific:

#### However, consider the following:

Species variations in pathogen proliferation rate may affect the amount and distribution of the pathogen in the body

### Challenges in test validation

 It is deemed useful and necessary to validate appropriate tests for a range of sample condition criteria such as different sample types, changes in detectability over time, under different storage temperatures, during autolysis, etc.



# Test constraints — Samples from wildlife

 Accessibility to diagnostic reference samples (adequate numbers, volume)

Positive reference samples pooling small quantities of samples from few infected individuals Dilution of a strongly positive sample to create a series of samples with different concentrations

Negative reference samples







# Test constraints — Samples from wildlife

### Quality of reference samples

- Opportunistic screening of dead animals is an effective way of monitoring wildlife populations for infectious agents
- a de Su de
- Disadvantage: compromised sample integrity (cross-contamination, autolysis)

- What to do?
- Ensure maximum utility of scarce samples
- Determine suitability for test validation, describe as (good, poor, autolysed)

# Test constraints — Samples from wildlife

**Only limited sample information available:** What is essential?

- a) the precise host species,
- b) specimen type



- c) geographical location with reference to known disease free or infected areas/regions,
- d) the date of sample collection
- e) Wherever possible, information on sex, age category (juvenile, sub-adult, adult), absence or presence of clinical signs, and a description of the signs will add value.



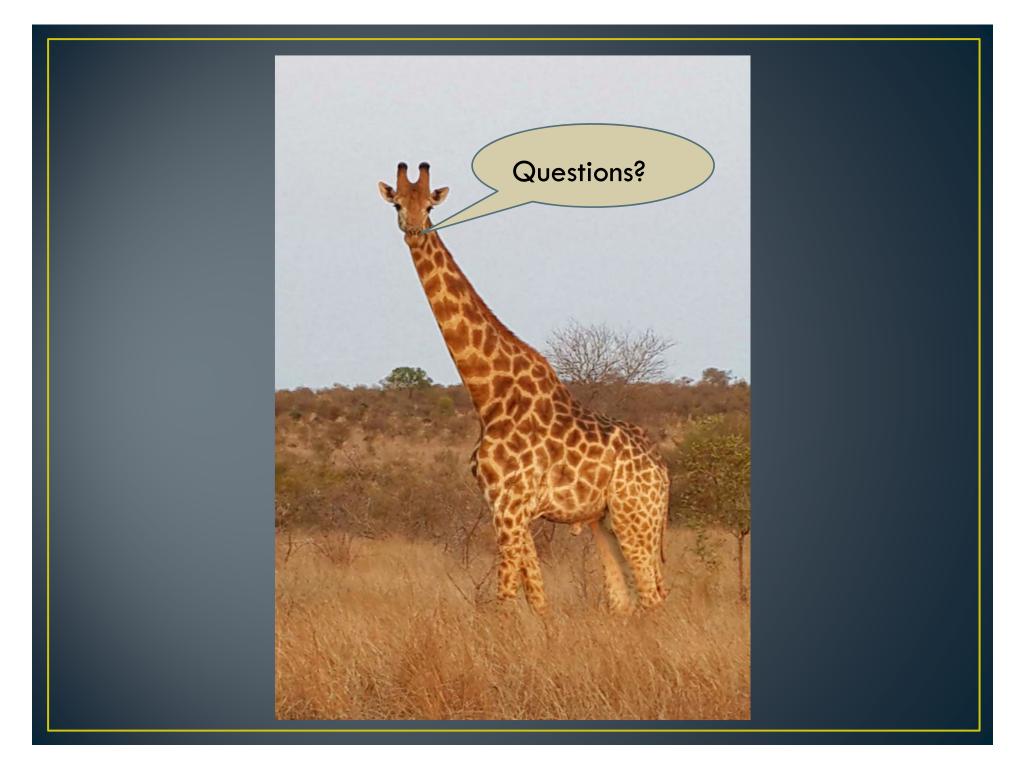
# In conclusion, ...

- Validation of diagnostic tests used in wildlife
  - has become important
  - has been made feasible due to the recommendation of the principle of provisional recognition

### However, ...

Non-validated tests can be worthwhile to use in a scientific approach (after all this is the way in which we improve existing methodologies)





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