2nd generation sequencing - a tool for exploring arbovirus distribution

Steve Kemp & Phil Toye, ILRI, Nairobi



AVID - Arbovirus Incident & Diversity

- icipe International Centre for Insect Physiology and Ecology
- KEMRI Kenya Medical Research Institute
- KARI Kenya Agricultural Research Institute
- KWS Kenya Wild life Service
- MoH Ministry of Health, DVBD, Kenya
- MPH Ministry of Public Health, Kenya
- DVS Department of Veterinary Services, Kenya

ILRI – International Livestock Research Institute



AVID - Arbovirus Incident & Diversity

ILRI's component -Sequencing and data management





THE UNIVERSITY of LIVERPOOL



The Wellcome Trust Funded Host-Pathogen Project





Questions

- "Where" is the virus (between "outbreaks") ?
 - Environment
 - Vectors
 - Reservoirs
- What is the diversity of ?
 - Virus
 - Vector
 - Reservoir
- And how do these interact ?
 - Distribution of other pathogens ?
 - Novel pathogens and variants ?

For example: Does a particular virus variant occur in a particular vector variant associated with a particular mammalian variant ?

Viral Geneflow



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Another complex mix of species, sub species & populations.

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Can we understand the dynamics of complex mixes of species, sub species & populations?

- (RT)-PCR Alone cannot do this.
 - It is a yes/no test (or a collection of yes/no tests).
 - It will not tell you about unknown sequences or sequence variants.
 - It will tell you about known variants but each requires its own PCR
 - It is very slow, laborious and time consuming when scaledup.

2nd Generation DNA sequencers

They are just DNA sequencers.

But their massively increased throughput means they provide a completely new way to do biology

They are tools for:

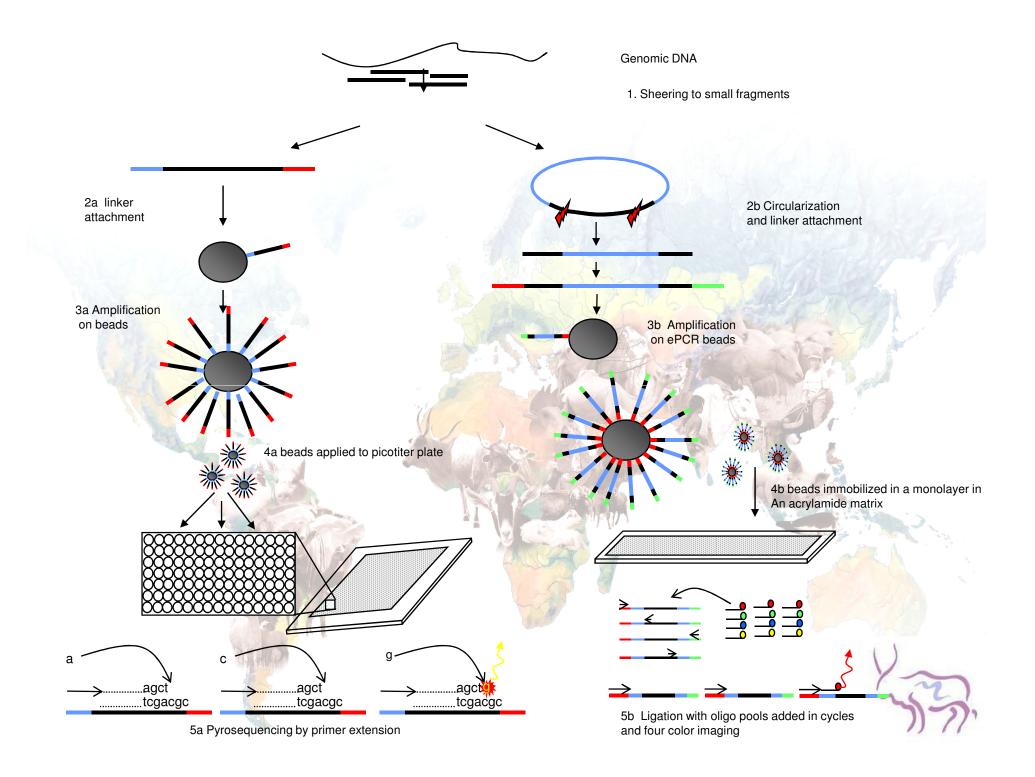
Probing complex biological mixes Uncovering diversity at species, sub-species, individual level

2 types

- 454 GSFLX
 - 500 Mbases in 7 hours
 - <u>£6,000</u> per run
 - 500bp reads

- ABI Solid
 - 30Gbases in 5 days (->90Gb)
 - £10,000 per run
 - 35 bp reads





2 broad approaches

1. Sequence everything in your sample

1. Reduce complexity by enrichment or amplification. Then sequence.

It all comes down to

- 1. how much complexity you can handle informatically and financially.
- 2. How much 'depth' you want.

2nd Generation DNA sequencers

Applications:

Metagenomics Genome diversity Genome function Digital expression analysis Pathogen discovery Epidemiology etc etc etc

Actually, this list is meaningless; applications run into each other.

Changing as technology improves !

- Sample potential vectors
- Sample people
- Sample wildlife

From 3 eco-climatic zones in kenya



- Each sample collected with a full meta data description (location, date/time, eco-geosocio descriptors)
- Simultaneously amplify and tag signature sequences from multiple points on multiple possible genomes – virus, insect, mammal, others.
- Sequence these *amplicons* simultaneously from 1,000s of samples.

- Analyse sequence look for distribution and co-occurrence.
- (Refine primers for a 'simple' (RT)-PCR approach.)

- This is already becoming obsolete. As capacity increases can we consider total sequence combined with sample fractionation? Eg low pass whole genome sequencing of all RNA viruses
- We can fully sequence approx.10 viruses on 1 machine run. [1st candidate - RVF vaccines?]



Outputs

• Information on diversity, dynamics, gene-flow, interaction to inform modeling and epidemiology

[Also ??

- A different kind of diagnostics ?????
 - Very high throughput
 - Not yes/no, but providing a signature for each virus
 - Can examine multiple pathogens simultaneously
 - Very cheap per sample, very expensive per run
 - NOT 'pen-side]
- NOT confounded by vaccination



When you have a hammer, every problem looks like a nail.







International Livestock Research Institute