Vaccine development for the control of African swine fever

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Disclaimer: None of the Vaccine opinions are from my previous employer at the USDA or my current employer Seek Labs, Inc. The opinions are solely my own based on published and collaborative studies.



USDA –NAVETCO Partnership 2020

African swine fever virus vaccine candidate ASFV-G-ΔI177L efficiently protects European and native pig breeds against circulating Vietnamese field strain

Manuel V. Borca and Douglas P. Gladue contributed equally to this study.

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- ¹ National Veterinary Joint Stock Company (NAVETCO), Ho Chi Minh City, Vietnam
- ² Agricultural Research Service | U.S. Department of Agriculture, Plum Island Animal Disease Center, Greenport, New York, USA

Days post challenge

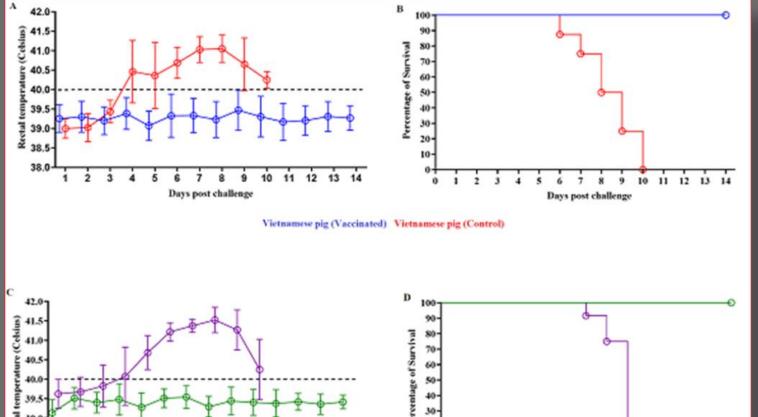
3 U.S. Department of Agriculture, Agricultural Research Service, Beltsville, Maryland, USA



7 8 9 10 11 12 13 14

Days post challenge





European pig (Vaccinated) European pig (control)



ASFV-G-ΔI177L is an effective vaccine in both Local breeds and European pigs



USDA, DAH, NCVI, NCVD- Additional I177L studies

No transmission in experimental conditions Limited transmission in field conditions (R0<1)

Overdose (10x dose) does not induce local or systemic problems

Field trials in North and South Vietnam confirm experimental safety data and 100% efficacy in representative group of animals challenged under experimental conditions

Reversion to virulence studies successfully completed: vaccine is phenotypically and genetically stable

Long term study (6 months) showed absence of residual virulence



ASFV Vaccine Development Status

Phase I Registration/ Phase III Licensing Phase II Discovery Vaccine Safety Regulatory Launch **Expanded trials** Field Trials Partnerships **Projects** and efficacy Approval 1. Vaccine for global market Involvement of PIADC scientists **ARS** patent CRADA.MTRA Vaccine for US-stockpile VÁC XIN NHƯỢC ĐỘC ĐÔNG KHÔ

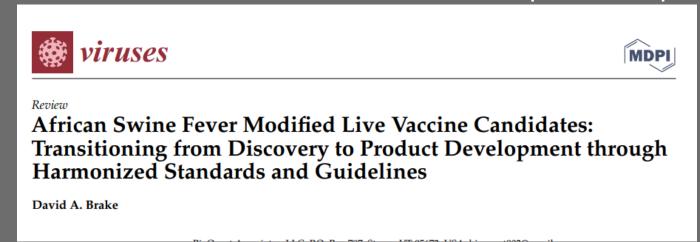
BỊCH TÁ BEO CHÁU PHI

ACCOUNTS THAT ASFV-G-ΔI177L – Navetco Licensing ASFV-G-Δ9GLΔUK (Undisclosed Partner) **AVAC ASF LIVE** ASFV-G-ΔMGF Aptimmune/AVAC ASFV-G-ΔI177L ΔLVR – Komipharma

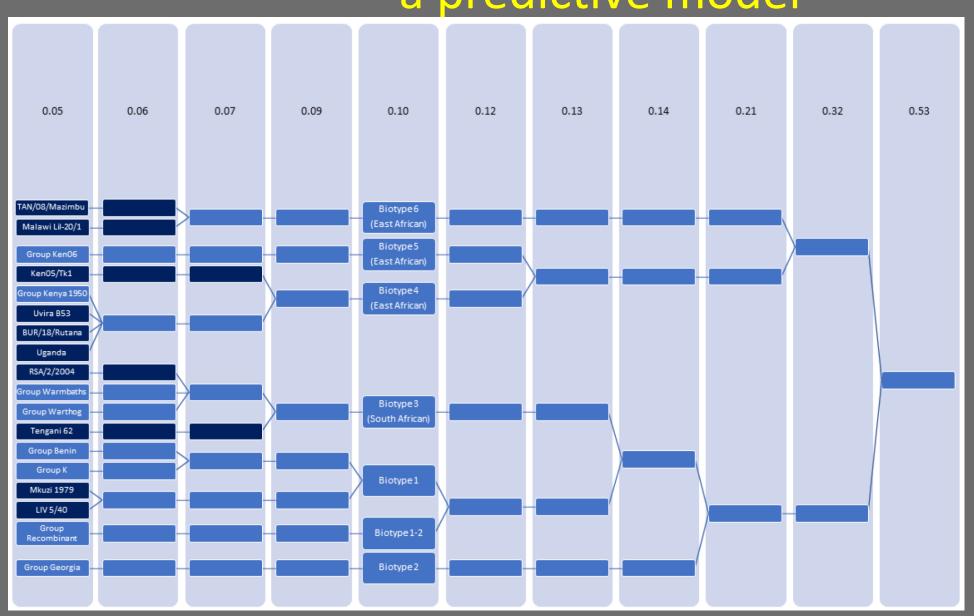
ASFV-G- ΔI177L ΔLVR-Dabaco

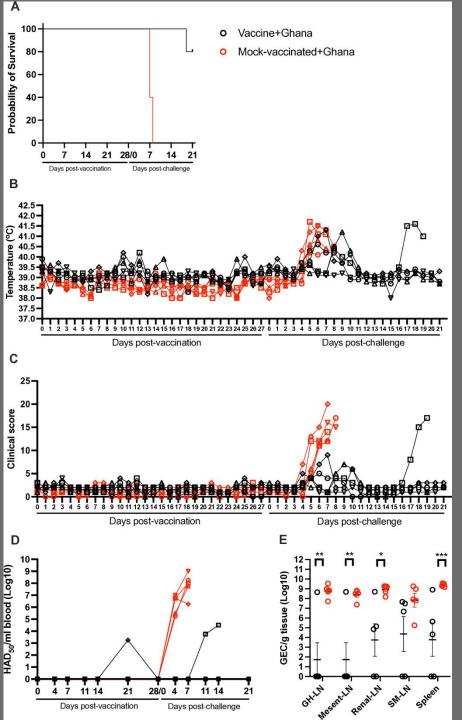
LAV ASFV Vaccines for widespread use

- Both Vaccines have been publicly under development for years
- LAV ASFV vaccines were first reported in 1996
- WOAH guidelines for safety studies for LAV vaccines are Still not available
 - The primary cause of delayed deployment of current vaccines
 - The cause for Random Reversion to Virulence studies with often misleading information, some pharmaceutical companies report on only on candidates they are not pursuing – destroy competition
 - David Brake as hired to interview ASFV experts and publish findings.



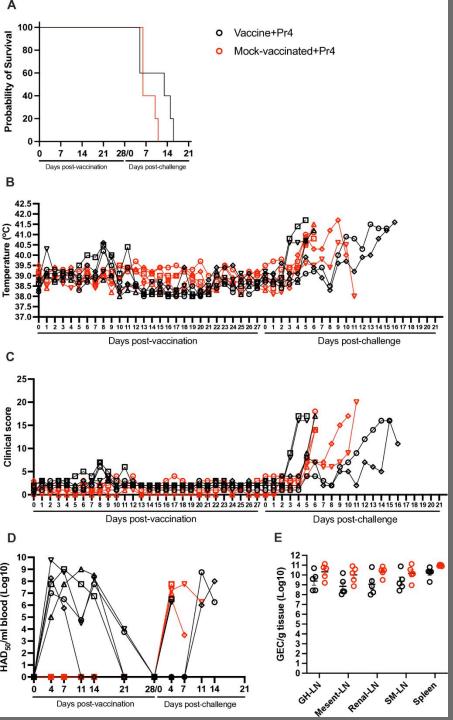
Cross-Protection of NAVETCO I177L using biotype as a predictive model





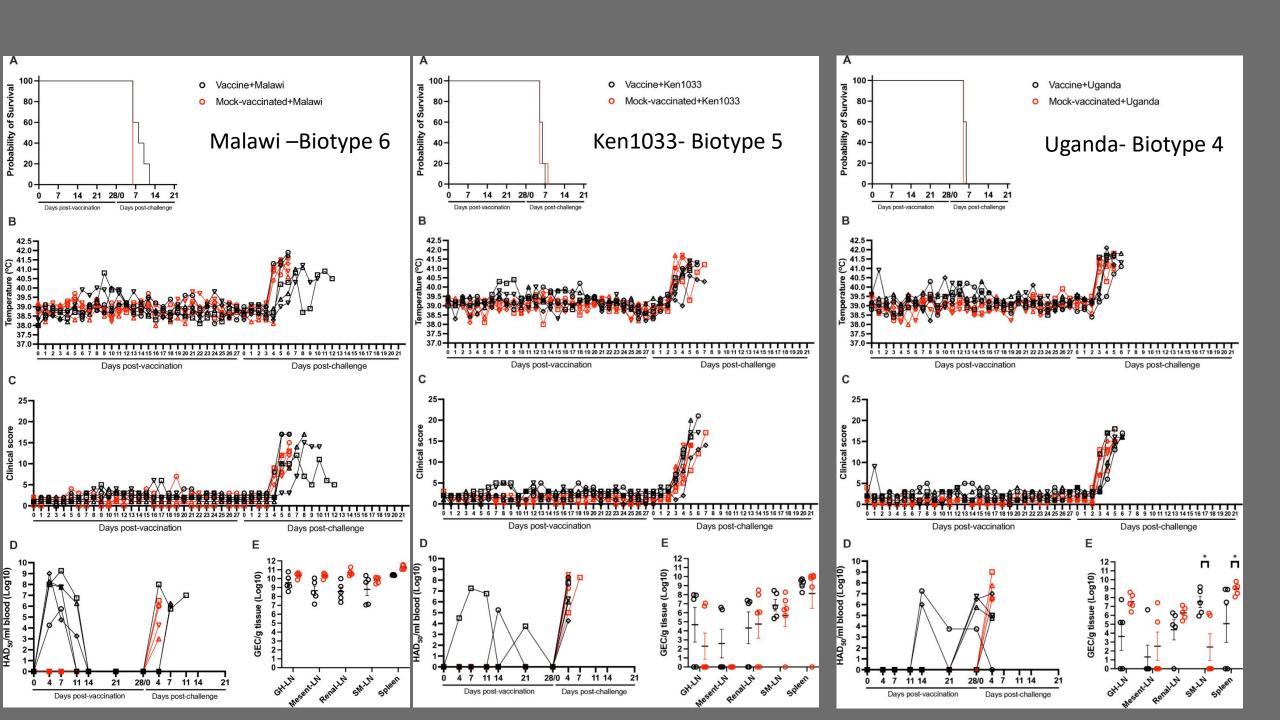
ASFV-G-ΔI177L Vaccinated

Ghana2021- Biotype I 80% survival

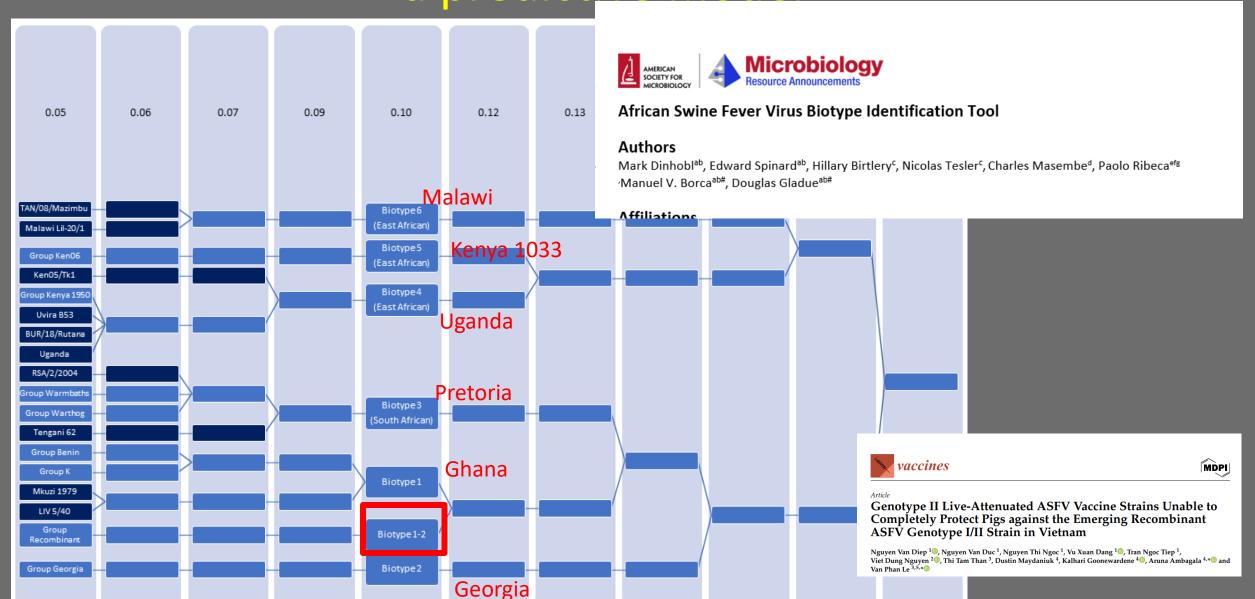


Pretoria Biotype 3 Identical P72 to ASFV-Georgia

Some extension of survival



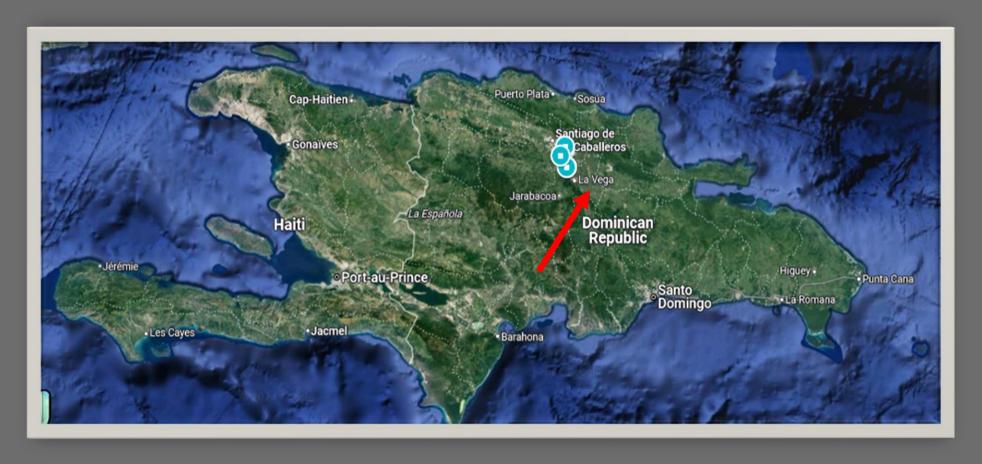
Cross-Protection of NAVETCO I177L using biotype as a predictive model



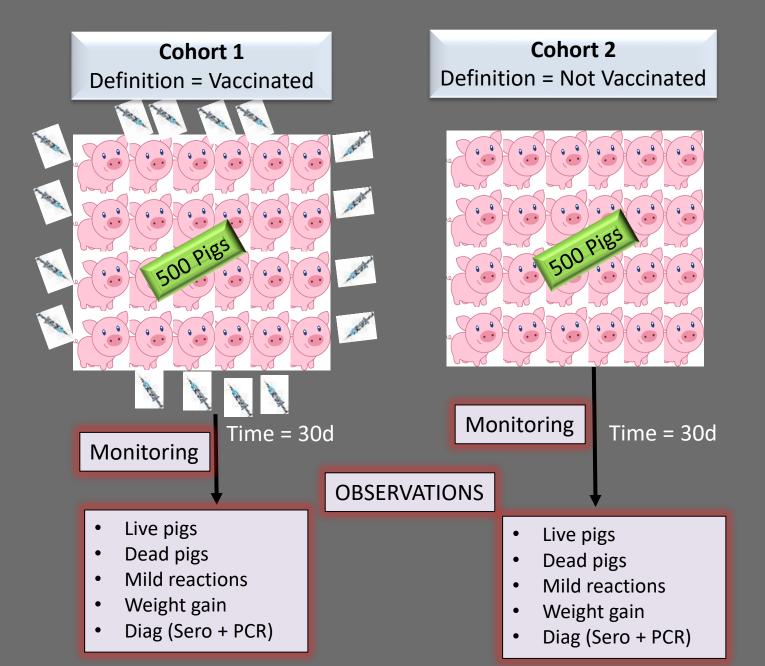
Genetic Differences to Predict Cross Protection?

- Biotypes are diverse too many changes to accurately predict which proteins could be involved for cross protection or lack of cross protection by Biotype Alone
- Why was there cross protection between Biotype II vaccines and Biotype I but not with Biotype II vaccines and Biotype I/II
- There were 74 proteins with genetic changes between Biotype I/II and Biotype II vaccine. 73 of these changes were also in Biotype I. Interestingly, the changes in Biotype I were additive to I/II.

Study Site: La Vega Province, DR

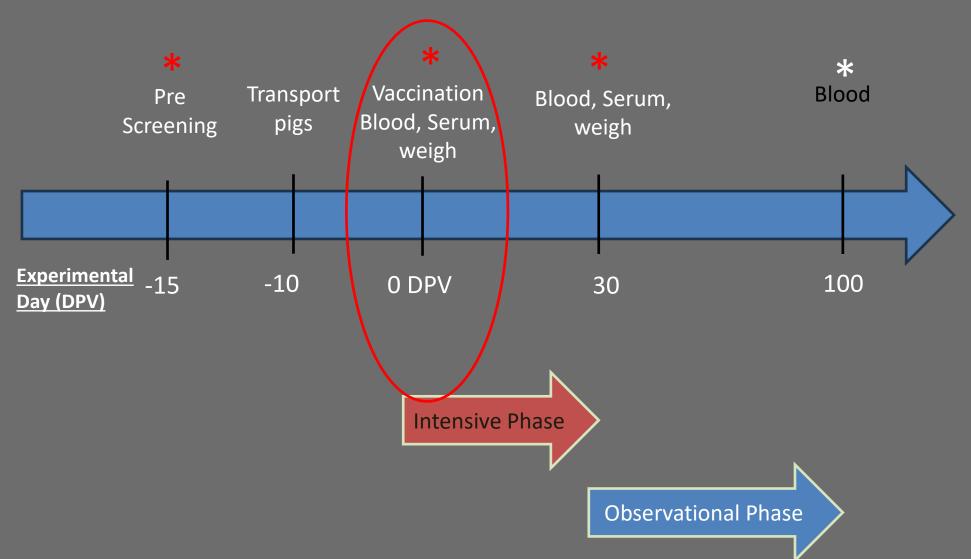


Prospective Cohort Study (Study Design)





Simplified study timeline (4 sampling points (*)



Results 3: Antemortem Diagnostics 0-30-90d

Purpose of samples

- Serum for anti-ASFV Ab (hint of efficacy)
- Blood for ASFV vacc DNA

Table 3. Serology for ASFV antibodies and detection of vaccine DNA in whole blood

	<u>Day 0</u>			
	ASFV ASF sero-			
	DNA	positive		
Vaccinated	0.0	0.0		
Control	0.0	0.0		

^{*}At day 100 blood was sampled for ASFV (vaccine) DNA, but not serology

Important points:

- Good seroconversion
- No transmission of vaccine
- Low prevalence and quantity of DNA detection in vaccinates

Results 4: fatalities

Total Since Vaccination	72 14.4%	56 11.2%	128
Total Fatalities	78	64	142
13-16	15	12	27
9-12	30	25	55
5-8	11	7	18
4	4	0	4
3	4	4	8
2	5	5	10
1	3	3	6
Pre-Vacc	6	8	14
Week Number	ì	, ,	
	(Vacc)	(Control)	per Week
	per Week	per Week	Total Fatality
	Fatality	Fatality	
	Total	Total	

Results 4b: Fatalities

	Traumatic Lameness		Hemorrhage (Mycotoxin) Syndrome		Neurologic Syndronie		Other Cause or Undetermined	
	Vacc	Control	Vacc	Control	Vacc	Control	Vacc	Control
Week Number								
(-1) Pre-Vacc	2	2	0	0	1	4	3	2
1	2	0	1	3	0	0	0	0
2	4	0	1	2	0	0	0	3
3	4	2	0	1	0	0	0	1
4	2	0	0	0	1	0	1	0
(5-8) Post- Experimental Observation Period		1	7	2	0	1	4	3
(9-12) Post- Experimental Observation Period	0	0	26	15	0	0	4	10
(13-16) Post- Experimental Observation Period	0	0	12	5	0	0	2	7
Total Since Vaccination	12	3	48	28	1	1	11	24

Shaded cell represents period in which category "Other" included cases consistent with PRRS

Conclusions (within this study):

- NAVET-ASFVAC did not cause differences in:
 - Temperatures of pigs
 - Weight gain
 - Fatalities

- Induced high prevalence of anti-ASFV antibodies
- Low prevalence and quantity of vaccine DNA at 90 DPV
- Vaccine was not transmitted between sheds when adequate biosecurity was implemented

What is the solution to ensure food safety with ASF spreading to secure the food supply?

- Subunit or Vectored vaccines? Likely not in the near future, likely require multiple antigens and are likely not cost effective.
- Live attenuated vaccines Safe when matched to outbreaks, and with biosecurity / government oversight
- Can we all agree that ASFV should be controlled?
- If no vaccines to control ASFV a universal therapeutic is urgently needed for food security.
 - Small molecules likely to produce disease resistant strains

 (as seen in HPAI, etc) when used widely on farms

A solution exists for ASFV and future emerging viruses without the need for RtV studies



Using CRISPR as a Broad-Spectrum Pan-genus Therapeutic To Rapidly Respond to Viral Threats: A Case Study for African Swine Fever and Beyond

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Programmable Target Ablation Platform

Seek Labs is transforming how diseases are treated by engineering CRISPR-based therapeutics using only sequencing information for rapid discovery, development, and deployment of precision therapeutics.



A CRISPR guide molecule is programmed to precisely target ands cleave pathogen's genome.



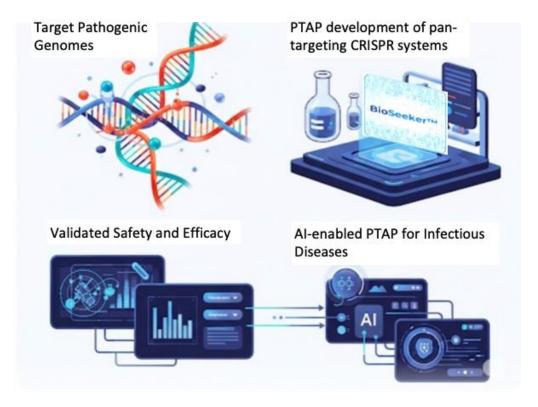
The invading pathogen is no longer able to replicate, limiting disease burden and spread to facilitate recovery.

Feature	RRET	Monoclonal Antibodies	Vaccines	Small Molecule Antivirals
Adaptability to Unknowns	Programmable per sample	Predefined antigens	Predefined viral genomes	Target-based discovery
Sample-to-Formulation	≤12 hours	3–6 months	6–18 months	Months to years
Per-Dose Latency	On-demand, ≤12 hrs	Batch-scale, weeks	Mass-manufactured	Dependent on production cycle
Manufacturing Requirement	In-system synthesis	Centralized, scaled	Biologic scale-up	Centralized synthesis
Delivery Infrastructure Required	Modular, point-of-care ready	Cold chain + infusion	Cold chain + global rollout	Oral or IV (centralized)



BioSeeker: Al-programmable Antiviral TX

BioSeeker automates and accelerates discovery and design of new therapeutics for Infectious Diseases



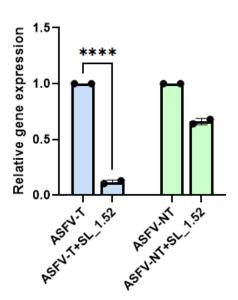
- Computational approach for Pan-targeting of Viral Diseases
- Disease dependent cleavage across all genomes
- Minimal number of gRNAs for efficient viral ablation
- gRNA design accounting for tolerated mismatches
- Structural tolerance of gRNAs for efficient Cas binding
- Processing features based on disease dependent positioning of multiplexed gRNAs
- Driven by constant updates on sequence and experimental data
- Proprietary machine learning algorithms for AIenabled design
- Delivery systems dependent on site or viral target
- Pre-screens gRNAs to ensure no off-targeting in host species



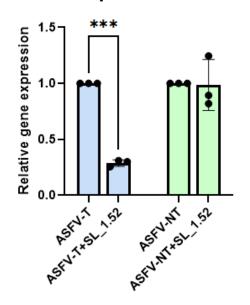
In Vitro Validation of ASFV Gene Knockdown

SL_1.52 Programmed CRISPR construct targets and cleaves a targeted ASFV gene (ASFV-T) in mammalian cells

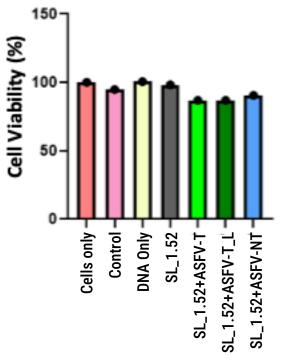
ASFV-T gene DNA reduced by 85%



Linked RFP reporter reduced by 75%



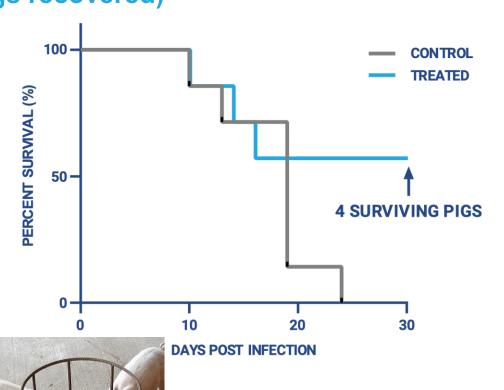
Minimal impact on cell viability



- SL_1.52 robustly and reproducibly knocks-down a targeted ASFV gene and
- SL_1.52 also decreased levels of a linked RFP-reporter gene in HEK293T
- Cell viability was not impacted by CRISPR activity

SL_1.52 In Vivo Efficacy: 4/7 pigs recovered from ASF

In one treatment cohort, 4 of 7 SL_1.52 treated ASFV-Infected pigs survived the trial (57% pigs recovered)

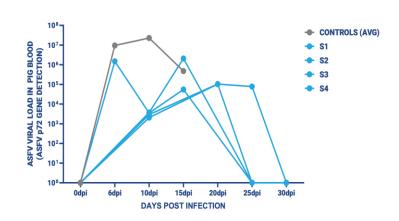


- Intramuscular lethal infection
- Treated on predicted days of clinical symptoms
- In ASFV-infected pigs, CRISPRtreatment enabled 4 of 7 pigs to fully recover from infection while all untreated controls succumbed to infection.
- Surviving pigs quickly recovered from initial clinical symptoms
- Natural challenge infection would be seen on infected farms, a less robust challenge test

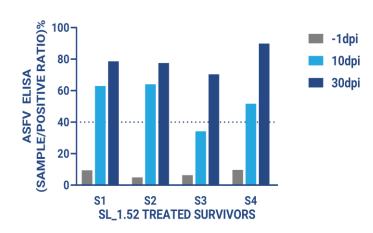
SL_1.52 Reduced Viremia and Increased Antibodies

Surviving pigs had lower ASFV viral load, robust antibodies and demonstrated clearance of virus in blood

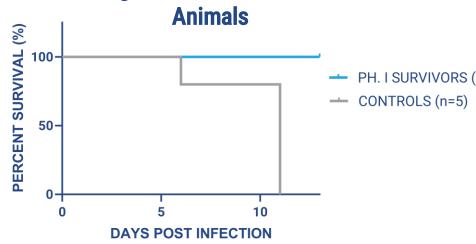
Treatment Muted Viral Replication in Survivors



Increased Pathogen-Specific Antibody Response in Survivors



Outcomes of Re-challenged Phase 1 Surviving Animals vs. Naive Control



- ASFV-CRISPR is an effective strategy to control the spread of ASFV
 - Has the potential for a universal treatment of ASFV
- First control approach that does not require strain-specific matching
 - Our CRISPR system targets conserved regions across multiple strains
- Our ASFV-CRISPR systems provide an inexpensive and accessible alternative to culling infected animals in outbreak or endemic areas.
 - In many endemic areas, governments do not have sufficient funds to reimburse farmers for loss resulting in quick sales of infected/exposed animals not showing clinical symptoms leading to further spreading of outbreaks to other farmers

Cross protection studies USDA-ILRI collaboration







Plum Island/NBAF

Elizabeth Ramirez Alyssa Valladares Leanna Burton Ediane Silva

Previous PIADC Involved in Presented vaccine work:

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Keith Berggren
Jolene Carlson
Paul Azzinaro
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Lauren Holinka-Patterson
Betty Bishop
Hillary Birtley

Mark Dihbol
Nino Vepkhvadze
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Navetco, Vietnam 2023

Thank you!

Questions?

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