

African Swine Fever Reference Laboratory Network



World Organisation for Animal Health Founded as OIE

FAO - WOAH Guidelines on ASF diagnostics and Pen-side tests

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Addressing ASF: Laboratory protocols and algorithms

- Published by the FAO in 2020.
- Basic PCR guideline with laboratory testing algorithm for ASFV
 - PCR protocol for King assay (WOAH-recommended)
 - Test worksheet
- Since then, lower virulence variants and new genotypes have emerged



Food and Agriculture Organization of the United Nations



Addressing African swine fever

Laboratory protocols and algorithms

INTRODUCTION

The Food and Agriculture Organization of the United Nations (FAO) and the World Organisation for Animal Health (IOE) including other partners have been working in countries affected or at risk of incursion by African swine fever (AS). This document was generated as guidance in response to the emergence of ASF in China, Southeast Asia, and the Pacific.

FAO has provided support for laboratory diagnosis of ASF following OIE recommendations, specifically using Polynemase Chain Reaction (PCR) in detecting ASF virus. PCR is a highly sensitive and specific method for the molecular detecting ASF virus for a wide range of purposes, including confirmation of clinical cases and confirmation of freedom from infection before movement. The Australian Center for Dissease Proparedness (ACDP, formerly the Australian Animal Health Laboratories) has developed a diagnostic algorithm based on OIE recommendations and in consultation with the Association of Southeast Asian Nations (ASEAN) regional animal health laboratory.

This document describes a validated real time reverse transcription-polymerase chain reaction (RT-PCR) protocol (the 'King assay'), which targets the *B646L* gene, encoding the ASF virus structural protein p72. This assay has been produced in kit from by the ACDP and provided to various veterinary diagnostic laboratories in Southeast Asia by the FAO and OIE. This document also provides links to other reference documents. FAO has provided three categories of guidance for the laboratory testing of pig samples for the presence of ASF virus:

- 1. Overview of primers and probe
- PCR protocols
 Surveillance laboratory flow char

1. Overview of primers and probes

Table 1. Primers and probes for the detection of ASF VIRUS in real-time PCR assays

ASF Assay	Forward Primer (5' → 3')	Reverse Primer (5' → 3')	Probe (5' → 3')	Dye	Que
King ⁴	CTIGCTCATIGGTATCAATCTTATCGA	GATACCACAAGATCRGCCGT	CCACGGGAGGAATACCAACCCAGTG	FAM	TAN
UPL	CCCASGRGATAAAATGACTG	CACTROTTCCCTCCACCGATA	GGCCAGGA!	FAM	Darl que
USDA	CCTCGGCGAGCGCTTTATCAC	GGAAACTCATTCACCAAATCCTT	CGATGCAAGCTTTAT	FAM	MG
McKillen	GTTGTTATGGAACSCGAAG	COCTCCTAGCTOGAAAGAAAA	CTIGAAAGTCCTCCGAGT	FAM	Edi Dari
Tignon	TGCTCATGGTATCAATCTTATCG	CCACTGGGTTGGTATTCCTC	TTCCATCAAAGTTCTGCAGCTCTT	FAM	TAN
Haines!	GATGATGATTACCTTYGCTTTGAA	TCTCTTGCTCTRGATACRTTAATATGA	CCACGGGAGGAATACCAACCCAGTG	da	0DC
Aguerolika	AGTTATGGGAAACCCGACCC	COCTGAATCEGAGCATOCT	NA	NA	NA

Emergence of naturally attenuated variants in China

- In 2020, virus isolates characterized as genotype II, with various changes in genomes (mutations, deletions, insertions, or shortfragment replacement)
- 11 isolates had non-haemadsorbing phenotype
 - Deletions or mutations in the EP402R gene (CD2v protein)
 - ▶ Two isolates tested *in vivo* had <u>lower virulence</u>, but highly transmissible
 - High dose: partially lethal (50-75%), caused acute or sub-acute ASF
 - Low dose: non-lethal, sub-acute or chronic disease, and persistent infection



Emergence and prevalence of naturally occurring lower virulent African swine fever viruses in domestic pigs in China in 2020

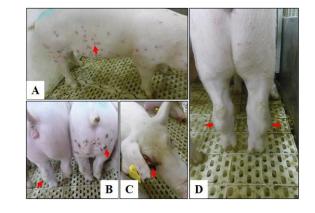
Sun Encheng¹, Zhang Zhenjiang¹, Wang Zilong², He Xijun², Zhang Xianfeng², Wang Lulu², Wang Wenqing¹, Huang Lianyu¹, Xi Fei¹, Huangfu Haoyue¹, Tsegay Ghebremedhin¹, Huo Hong¹, Sun Jianhong¹, Tian Zhijun¹, Xia Wei¹, Yu Xuewu³, Li Fang², Liu Renqiang², Guan Yuntao¹, Zhao Dongming⁴ and Bu Zhigao⁵



Emergence of Genotype I in China

ORIGINAL ARTICLE

- In 2021, detection of Genotype 1 viruses for the first time
 - High levels of genomic similarity to historical isolates from Portugal
 - Non-HAD phenotype, with mutations in the EP402R gene (CD2v)
 - Low virulence, efficient transmissibility
- Caused mild infection and chronic disease
 - Paralysis, weight loss, intermittent fever, skin ulcers, arthritis and sporadic deaths, swollen joints, papules on the skin and necrotic lesions



Genotype I African swine fever viruses emerged in domestic pigs in China and caused chronic infection

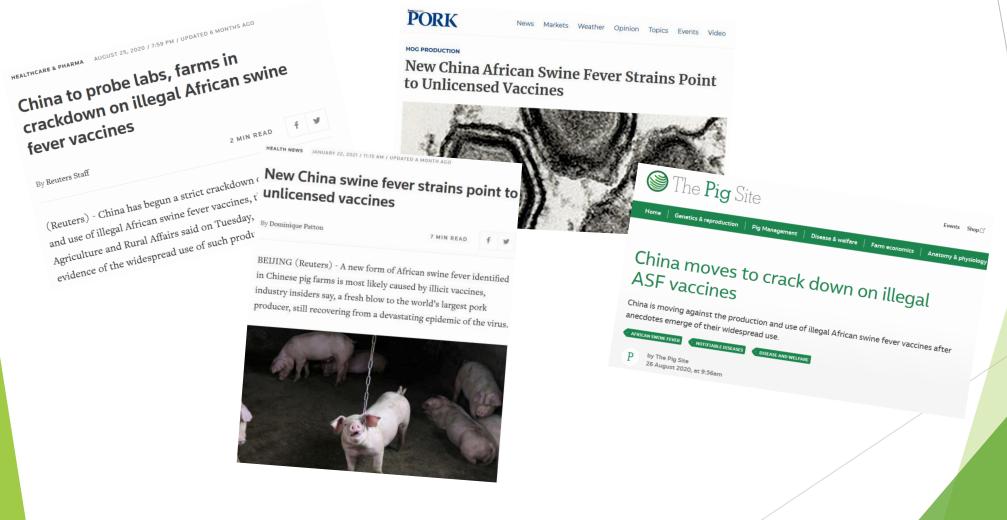
Encheng Sun*, Lianyu Huang*, Xianfeng Zhang*, Jiwen Zhang*, Dongdong Shen*, Zhenjiang Zhang, Zilong Wang, Hong Huo, Wenging Wang, Haoyue Huangfu, Wan Wang, Fang Li, Rengiang Liu, Jianhong Sun, Zhijun Tian, Wei Xia, Yuntao Guan, Xijun He, Yuanmao Zhu, Dongming Zhao © and Zhigao Bu 💿

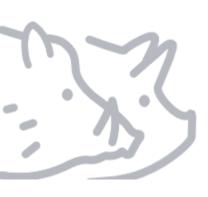
State Key Laboratory of Veterinary Biotechnology, National High Containment Facilities for Animal Diseases Control and Prevention, National African Swine Fever Para-reference Laboratory, Harbin Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Harbin, People's Republic of China



fever vaccines

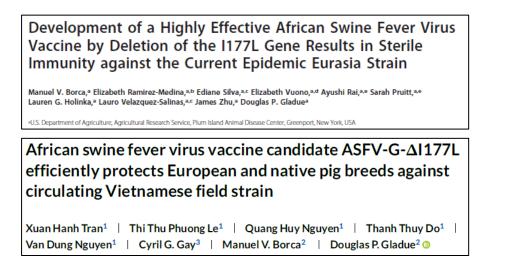
By Reuters Staff

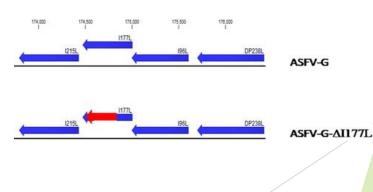




Vaccine progress: USDA/NAVETCO LAV

- Developed by USDA
 - Recombinant Live attenuated vaccine candidate based on Georgia/07 (G2)
 - Deletion of previously uncharacterised gene (I177R)
 - Protects against virulent ASFV challenge in experimental studies





Updating the ASF Diagnostic Protocols and Algorithm

- With new variants emerging and novel vaccines undergoing trials in Vietnam and China, an updated laboratory guideline is needed
- A new version of the ASF Manual was developed based on WOAH recommendations for laboratory diagnosis of ASF
 - Address detection of virulent and variant forms of ASF and the potential future use of authorised LAVs with defined mutations
 - ► Provide guidance for early detection of ASF infection by viruses causing different clinical forms of disease (e.g. acute, subacute, chronic and subclinical) → requires both <u>PCR</u> and <u>serological</u> diagnostic testing for sensitive and accurate lab diagnosis



Updates to PCR protocols

- Assays selected:
 - Detection of G2 variants with deletions in the MGF360/505 and EP402R genes, associated with sub-clinical and chronic ASF
 - Differential detection of G2 LAV-derived virus containing a deletion in the I177L gene
 - Detection of G1 viruses associated with lower virulence and chronic disease
- All assays have been validated (published or by ASF Reference laboratories)



Triplex PCR assay

- Developed by the Chinese Animal Health and Epidemiology Center for multiplex detection of MGF360/505, EP402R (CD2v) and B646L (p72) genes (Dr Zhiliang Wang)
- DeteValidated using Chinese ASFV isolates belonging to genotype II and MGF360/505 and EP402R (CD2v) genes deletion LAV candidate strains developed by CAHEC
- ct and identify variants containing genome deletions in the MGF360/505 and EP402R (CD2v) genes
- Investigation of subacute and chronic ASF

	Test results			
Interpretation	P72- FAM	CD2v- VIC	MGF-Cy5	
ASFV pandemic strain positive	+	+	+	
ASFV CD2v gene deletion strain positive	+	-	+	
ASFV MGF gene deletion strain positive	+	+	-	
ASFV CD2v and MGF gene double deletion strain positive	+	-	-	
ASFV negative	-	-	-	

Note: "+" means the test is positive; "-" means the test is negative.



ASFV-G-ΔI177L LAV PCR assay

- Developed and validated by the United States Department of Agriculture (USDA), Agricultural Research Service, at the Plum Island Animal Disease Center (Velazquez-Salinas et al., 2021. Oct 27;8:768869)
- ► Genetic DIVA test for the specific detection of the ASFV-G-∆I177L LAV candidate
- Use for further molecular characterisation of samples that have tested positive for p72 detection using either the King or the Triplex assay

Development of a Highly Effective African Swine Fever Virus Vaccine by Deletion of the I177L Gene Results in Sterile Immunity against the Current Epidemic Eurasia Strain

Manuel V. Borca,^a Elizabeth Ramirez-Medina,^{a,b} Ediane Silva,^{a,c} Elizabeth Vuono,^{a,d} Ayushi Rai,^{a,a} Sarah Pruitt,^{a,a} Lauren G. Holinka,^a Lauro Velazquez-Salinas,^{a,c} James Zhu,^a Douglas P. Gladue^a

«U.S. Department of Agriculture, Agricultural Research Service, Plum Island Animal Disease Center, Greenport, New York, USA



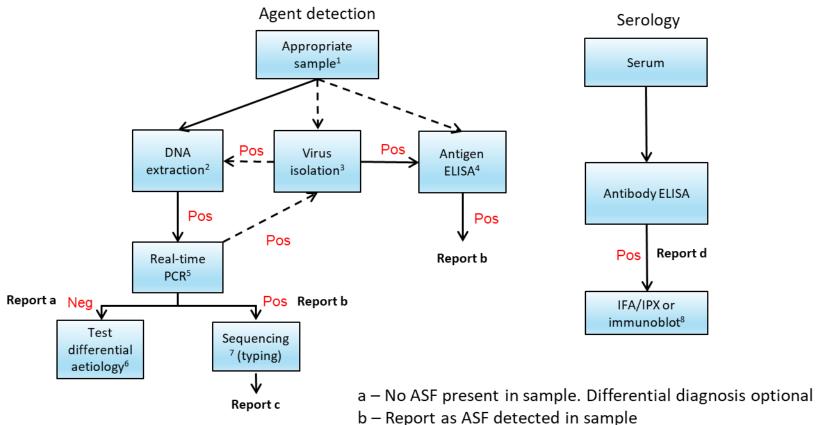
ACDP ASFV Genotype 1 PCR assay

- ► Targets 505-3R gene
- Validated using a panel of reference isolates belonging to genotypes 1, 2, 7, 9 and 10, as well as diagnostic specimens
- Should be used for:
 - Investigation of subacute and chronic ASF
 - Further molecular characterisation of samples that have tested positive for p72 detection using either the King or the Triplex assay





Laboratory algorithm



- c Report as ASF detected in sample with genotype/types
- d ASF antibodies detected in sample



ASF Point of Care testing guide

- Several diagnostic platforms available commercially
- The WOAH ASF Reference Laboratory Network has recently drafted an overview of commercially available tests
 - Technical details, costs, advantages vs disadvantages
 - Based on peer-reviewed publications or independent evaluation at Ref lab
 - For ASF diagnosticians, field workers and decision makers





The OIE ASF Reference Laboratory Network's overview of African swine fever diagnostic tests for field application



February 2022



ASF Point of Care testing guide

- Guide is available in English and Spanish, and is being translated into Japanese, Chinese (Mandarin), Bahasa Indonesia, Vietnamese, and Thai
- Free download from:

https://rr-asia.woah.org/en/news/the-oie-asfreference-laboratory-networks-overview-ofafrican-swine-fever-diagnostic-tests-for-fieldapplication/



The OIE ASF Reference Laboratory Network's overview of African swine fever diagnostic tests for field application



ASF Rapid Tests

Detects virus antigen or antibody in blood samples from infected pigs. Can be used for disease investigations, as part of ASF surveillance.

Advantages:

- Use in the field, pen-side
- Simple to use
- Convenient
- Inexpensive
- No instrumentation
- Small volume of sample
- Highly specific

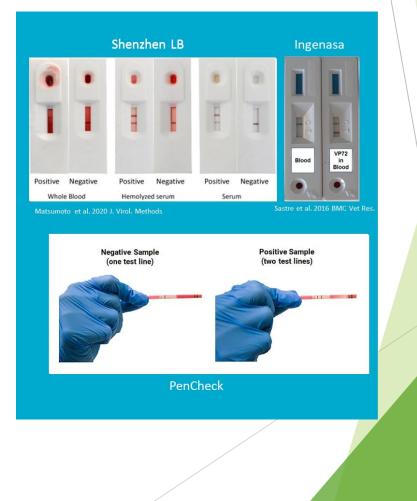
Disadvantages:

- Lower diagnostic sensitivity (Antigen)
 - 60-70%
- Prone to false-negatives
- Can only be used with un-clotted whole blood (Antigen)



Field tests: Antigen detection

- Several commercial options
 - Lateral flow or dip stick
 - ▶ Rapid (15-30 mins) and simple to use
 - ► Inexpensive (~\$USD 3-14)
- Typically less sensitive than molecular tests, but can have comparable specificity
- Recommended to:
 - be used on sick or dying pigs (higher levels of viraemia)
 - test samples from more than one sick pig to increase the chances of detecting infection





Field tests: Molecular Tests

- PCR or isothermal methods
- Sensitivity and specificity comparable to lab-based real-time PCR

→ detection of ASFV infection (viral DNA) at early stages

- 0.5-2 hours
- Also used for detection of contaminated carcases, pork and environmental samples at point-of-need (e.g. abattoir, airport, wild boar/feral pig habitats)
- Technically more complex, require much higher level of <u>training</u> and QC/QA, and <u>expensive equipment</u>



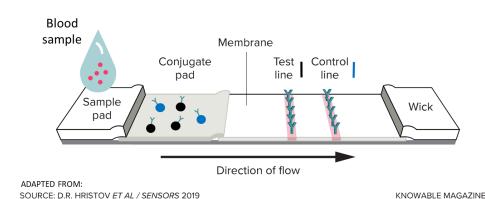


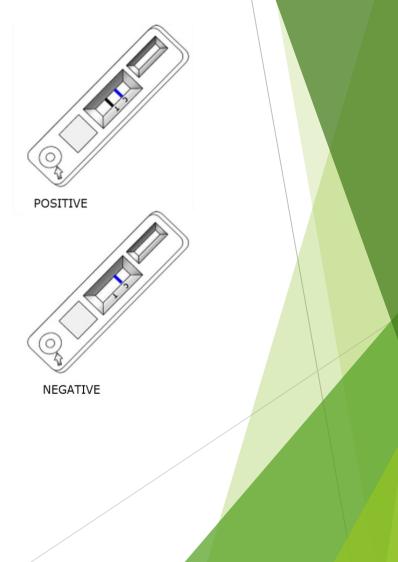
Source: Dr. Ken Inui, FAO



Field tests: Antibody Tests

- Lateral flow devices
 - Rapid (15-30 mins) and simple to use
 - Inexpensive (~\$USD 5-17)
- Immunochromatographic test
- Comparable diagnostic performance to ELISA
 - ► Specificity ~100%
 - Sensitivity lower than IPX/IFA

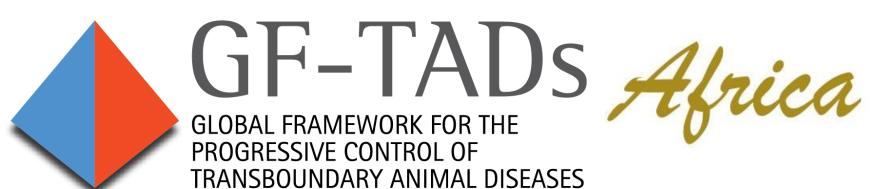






What to choose?

- Influenced by many factors including costs, ease of use, training requirements, sample type to be tested
- Simple rapid tests may be appropriate for certain situations, such as resource-poor settings
- More advanced molecular platforms may be the test of choice in settings where costs are not a major factor and operators can be confidently trained to a high level of competency
- For some countries, a combination of tests may be employed depending on application and available resources





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