

EARLY DISEASE DETECTION

SAMPLING AND DIAGNOSTIC TESTS IN APPARENTLY HEALTHY POPULATIONS

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Traditional approaches to Diagnosis & Prognosis

- How do we detect abnormalities?
- Experience / training, gut reaction & intuition
- Some wise individuals just know from experience (and ability to convince others that they are correct)
- How do we translate diagnosis in sick animals to surveillance in apparently healthy animals?

Why use apparently healthy animals

- Preventive medicine depends on early detection
- Early detection cannot wait for mortalities to start increasing
- So using healthy population, maybe healthy individuals to satisfy need for earlier detection

Sampling principles

- Random sampling (= probability sampling)
 - Simple random
 - Systematic random
 - Multiple staged, random sampling
 - E.g. area / farm / pond
- Biased sampling
 - Convenience
 - Risk-based = syndromic

Random Sampling

- Systematic random sampling is most common form of random
 - Want 100 samples from 10,000 fish unit
 - Then take every 100th fish, starting with random in first 1-100
 - E.g. 27, 127, 227,, 10027, 10127
- Can apply to different levels of concern



Biased sampling

- Convenience samples
 - Feeding and catching fish with dip net



Biased sampling

- Risk-based (i.e. syndromic) samples
 - Moribund with specific external characteristics known to be more common for disease of interest



Disease detection

- Diagnostic tests are imperfect
 - Particularly when attempting to detect asymptomatic individuals
- ▶ New cultured species will have new pathogens identified

Diagnostic tests in aquatic situations

- Disease is frequently assumed to be absent in facilities that lack obvious clinical disease
- Subclinical and carrier states are a major consequence for non-stressed animals
- Diagnostic tests cannot detect many non-clinical disease states
 - Many new diseases have no tests except general pathology to identify suspicious agents or host responses

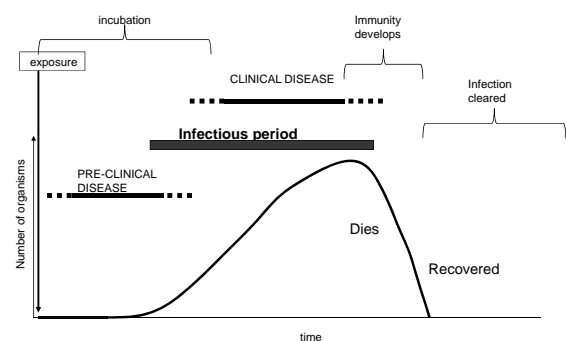
Comment about working with new species

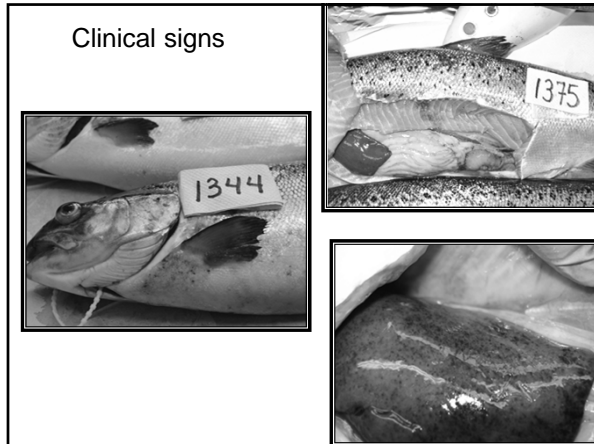
- New diseases (i.e. new host-pathogen interactions) are most likely to occur when working with new species AND working with several new species at same time
- Not likely to recognize the event when it first happens

Virus X example

- Diagnostic tests (ELISA)
 - Correctly detect 96% of positive cases (sensitivity)
 - Correctly detect 98% of negative cases (specificity)
- So why do we approach a positive test done in a healthy animal much differently than a positive test in a clinically sick case?

Pathogen replication in host





Disease Surveillance

- Sample from 25,000 fish cage
 - From 10-20 cages per site
 - From 75-90 sites
- Test with which test
 - Virus isolation - 3-21 days, \$100/test
 - Indirect fluorescent antibody test (IFAT) – 1 day, \$15/test
 - Enzyme Linked Immunosorbent Assay (ELISA) – 1 day, \$20/test
 - RT-PCR – 1 day, \$45/test

Background

- Sensitivity (Se): Probability that an infected fish will test positive
- Example: Se = 62%

	D+	D-
T+	62	
T-	38	
	100 fish	

Background

- Specificity (Sp): Probability that an uninfected fish will test negative
- Example: Sp = 98%

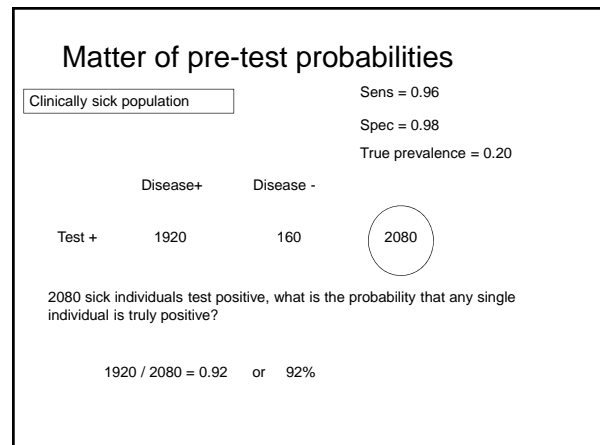
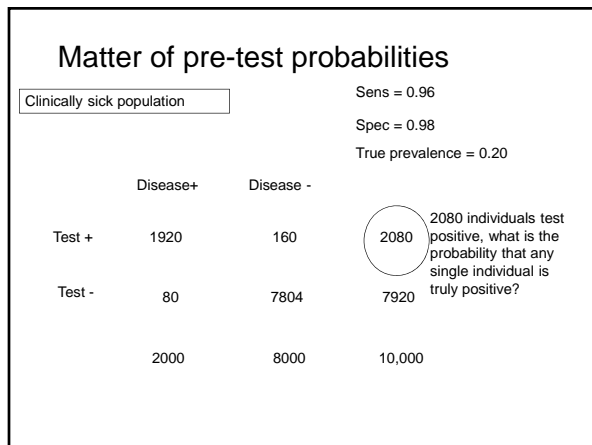
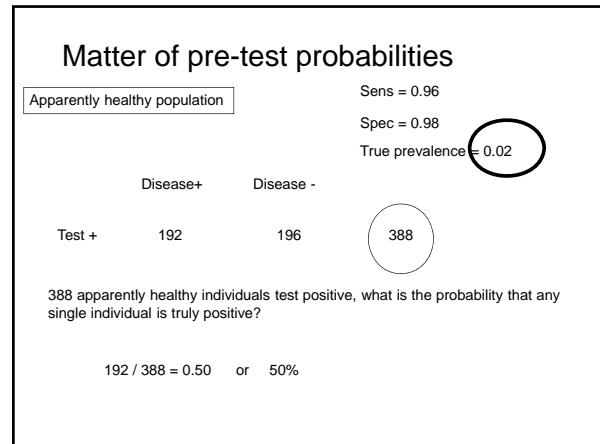
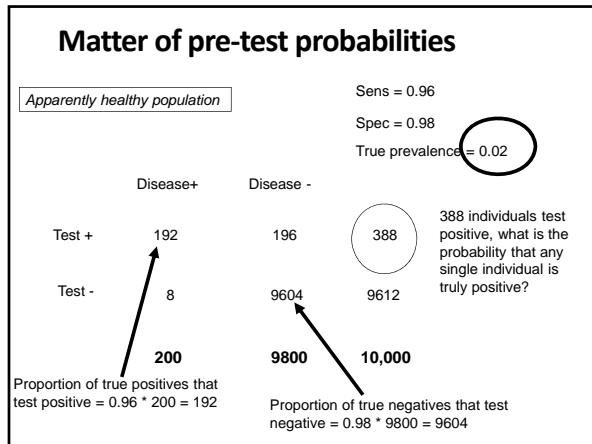
	D+	D-
T+		2
T-		98
		100 fish

Background

	D+	D-	
T+	62	2	64 positive tests
T-	38	98	136 negative tests
	100 fish	100 fish	200 total tests

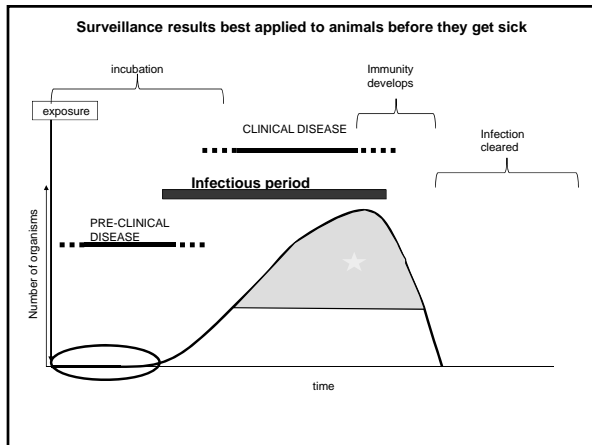
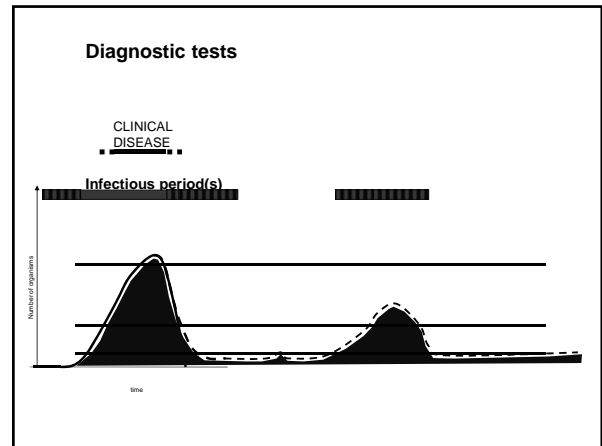
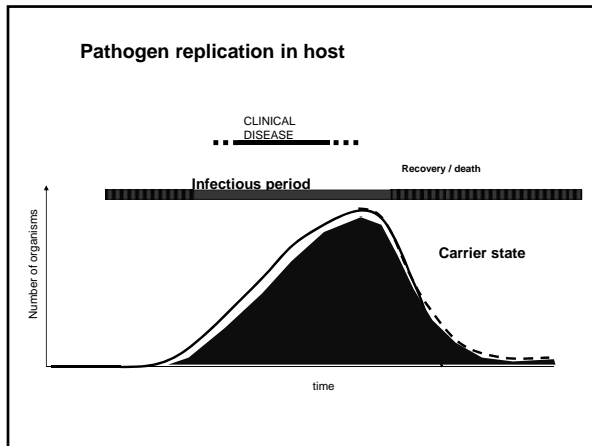
Biasing samples can be good

- We routinely bias our samples toward detection
 - By looking for individuals that have characteristics common in the diseased population
 - Smaller individuals (compared to cohorts)
 - Off-feed or altered swimming behaviour
 - Slow swimmers
 - Fish with lesions



- Experienced clinicians are good at increasing the prevalence of a condition through identification of clinical signs (physical exam) and history
- diagnostic tests then have a higher probability of being correct
- Note: physical exam for clinical signs can be assessed for sensitivity and specificity because they are subjective "diagnostic tests"

- ### Pathogens: The Host Perspective
- Clinical disease
 - Externally obvious abnormality usually leading to decreased probability of surviving or reproducing
 - Direct result of abnormality or indirect (e.g. increased predation)
 - Subclinical disease
 - Abnormality is not externally obvious (behaviour or lesions absent), but change in probabilities likely
 - Usually reflects our reliance on diagnostic tests for agent or host response to diagnose



- Detection is affected by many factors**
- Disease level in the individual tested
 - Clinical disease is easier to detect
 - Surveillance of 'apparently healthy' individuals is more difficult
 - Disease level in the population
 - Which animals are sampled

Depopulation of cage: Does it prevent virus exposure for other cages at site?

Apparent prevalence of different populations (HPR_{ALL}) of Atlantic salmon in New Brunswick farms (2001).

Population	Apparent prevalence (95%CI)	
A ^a	0.940 (0.887, 0.993)	moribund fish, outbreak cage
B ^β	0.406 (0.279, 0.533)	healthy fish, outbreak cage
C ^β	0.286 (0.204, 0.368)	healthy fish, non-outbreak cage, outbreak site
D ^γ	0.084 (0.009, 0.160)	healthy fish, non-outbreak cage, non-outbreak neighbor site
E ^γ	0.080 (0.004, 0.156)	healthy fish, non-outbreak cage, distant site

McClure, Hammell, Dohoo, Nerette, Hawkins. 2004. J Fish Dis 27: 375-383.

- Diagnostic test performance**
- What happens when an individual is sick with a virus?
 - When does infection actually occur?
 - When will clinical signs be evident?
 - When does a diagnostic test work best?

Conclusion

- Diagnostic samples must occur in surveillance program
 - Collecting evidence to change our decisions about what is positive and what is negative when fish are sampled *prior* to actual mortality spikes
- Depopulation may be only control measure
 - Costs are high so don't want to decide too early
 - Allowing positive cases to continue can cause entire site and area to continue positive (cost much more!)