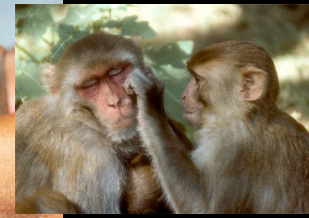
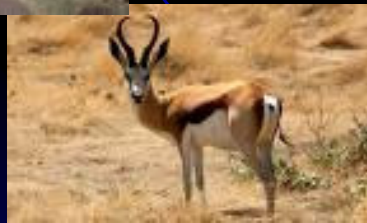


# The Impact of wildlife in the epidemiology of RVF

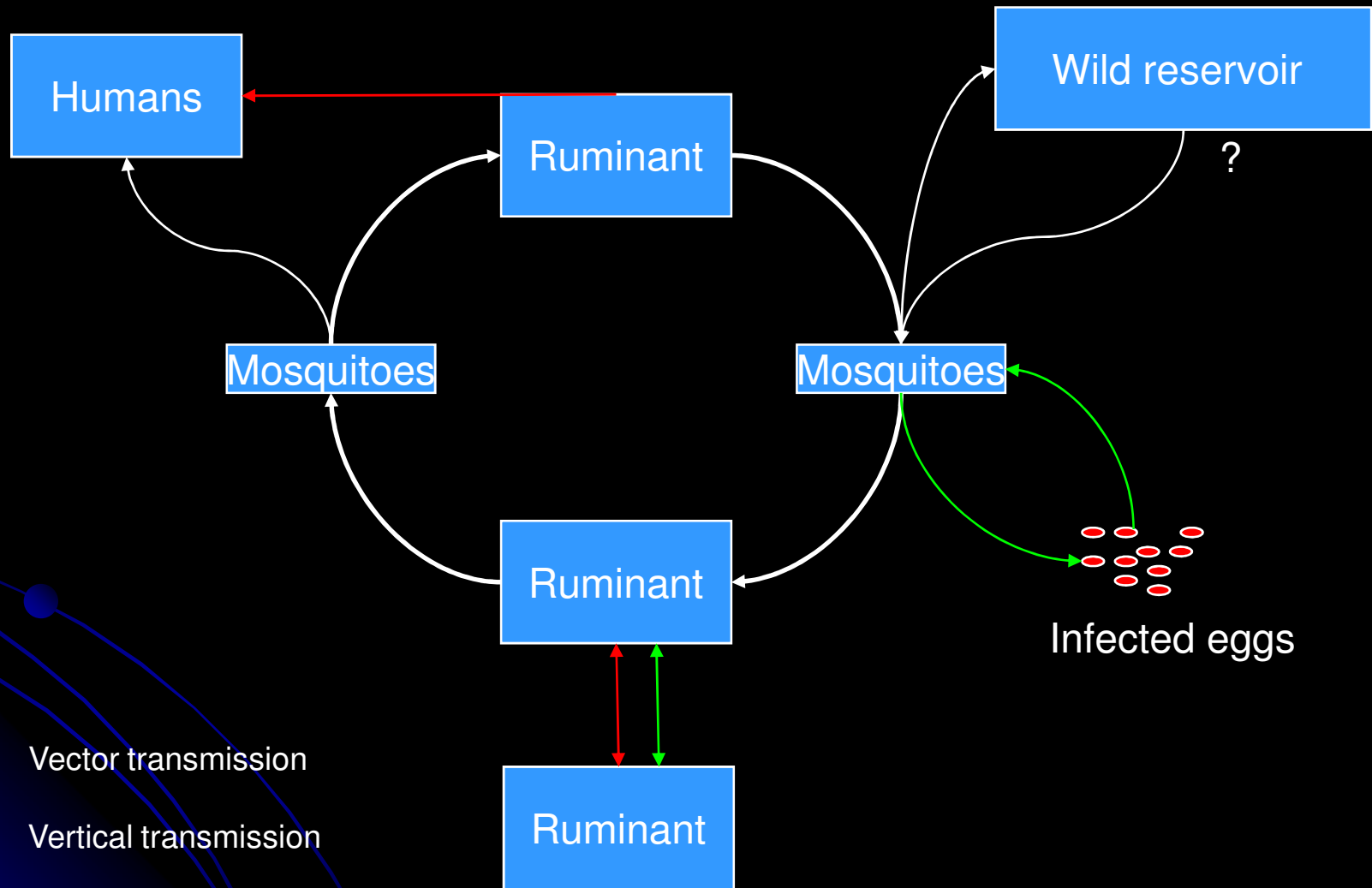
**V. Chevalier**

UR AGIRs « Animal et Gestion  
Intégrée des Risques »  
CIRAD – ES



# Introduction

## Theoretical epidemiological cycle of RVF



- Vector transmission
- Vertical transmission
- Direct transmission

# What is a reservoir ?

- Many different and often contradictory definitions

(Haydon et.al, EID, 2002)

- Pathogenic (WN and birds in US) / non pathogenic (WN and birds in EU)
- 1 host (badger and M. Bovis in UK) / multi-host system (dog+jackal and rabie in Zim)

- Indicators to identify reservoirs

- Evidence of associations
- Quantitative data on risk factors: case control studies
- Identifying natural infection : antibodies, virus
- Experimental inoculation
- Persistence of the virus in the reservoir population = longitudinal studies



# Litterature review

## Rodents

- Inoculation on *Apodemus sylvaticus*, *Microtus agrestis*, *Muscardinus avellanarius* (Daubney, 1931)  
=> suspected of being reservoir
- Heavy mortality in *Arvicanthis abyssinicus* and *Rattus rattus* in an infected farm (Kenya, 1932)
- Wild field rat, *Arvicanthis Abyssinicus*, able to be infected by RVFV, developed viraemia without succumbing to the infection (Weinbren , Uganda, 1957),

# Rodents : serological evidence



Ref year	Species	Location	Method	Confirmation test	%
1978	<i>A.Niloticus</i> <i>A.Cahirinus</i> <b><i>Rattus rattus</i></b>	Egypt	HI	No	21.5 8.2 3.1
1982	<i>Gerbillus sp</i> <i>Meriones crassus</i>	Egypt	HI	PRN	19 5
1983	<i>Mastomys</i>	RCA	IFI	TDRP	3.6
1987	<i>Mastomys erythroleucus</i>	Senegal Mauritania	IFI	No	0.75 (n= 57)
1997	<i>Aethomys namaquensis</i>	South Africa	ELISA	NT	15 (n=312)
2000	<b><i>Rattus rattus</i></b> <i>Mastomys huberti</i> <i>A.Niloticus</i> <i>M. erythroleucus</i>	Senegal	NT	ID	50 13;5 4;3 2;4
2001	<b><i>Rattus rattus</i></b>	Egypt	ELISA	No	29.3 (n= 300)



# Rodents : virological evidence

Year	Location	Species	Test	% positive
1979 *	Egypt	<i>Rattus Rattus</i>	CF	12.5 (n=8)
2001 **	Egypt	<i>Rattus Rattus</i>	RT-PCR	9.6 (n=300)

- Imam et al, 1979
- \*\* Youssef and Donia, 2001



# Other species



- Antibodies in wild ruminants in Zimbabwe (IHA confirmed by ELISA)
  - African buffaloes (*Syncerus caffer*) 6.2% (n=541)
  - Waterbuck (*Kobus ellipsiprymnus*) 4.5% (n=179)
  - White rhino (*Ceratotherium simum*) 8.3% (n=84)
  - Black rhino (*Diceros bicornis*) 16% (n=110)
- Antibodies in African buffaloes in Kenya (1998) (Davis, 1975)
- Viraemia after experimental inoculation:
  - African buffaloes (*Syncerus caffer*) (Kenya, Daubney 1932) (Davies, 1981)
- Abortions in South Africa (Joubert, 1951)
  - Springboks (*Antidorcas marsupialis*)
  - Damaliscus (*Damaliscus albifrons*)
- Presence of antibodies in hippopotami and elephant, and abortions in springbok and blesbok during epidemics. (Bengis and Erasmus (1988))

# Other species



- Bats

- Two viral strains isolated from *Micropteropus pusillus* and *Hipposideros abae*, with positive serological test on humans and ruminants (, Boiro *et al.* (1987)) , Guinea
- Isolation from organ pools of *Micropteropus pusillus*, *Miniopterus schreibersi*, and *Hipposideros caffer* (Konstantnov *et al.* (2006)), Guinea

- Monkeys

- Serosurvey on 333 baboons Kenya (Davies *et al.*, 1971)
- Experimental inoculation => viraemia and antibodies)(Nilklasson *et al.*, 1983)

- African carnivores species

- Warthog



# Geographical distribution and persistence mechanisms

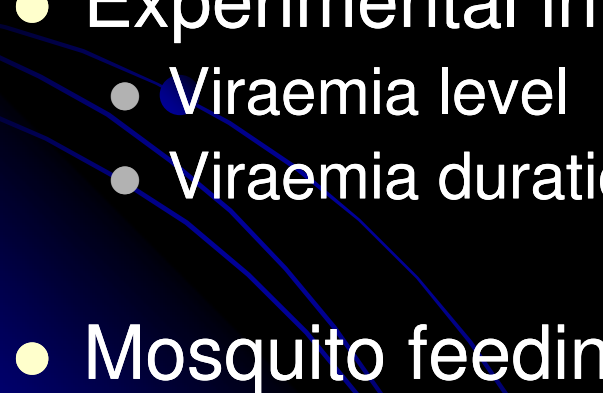


Enzootic circulation and/ or outbreaks



Sporadic cases and/or viral isolations and/or infection serological evidence

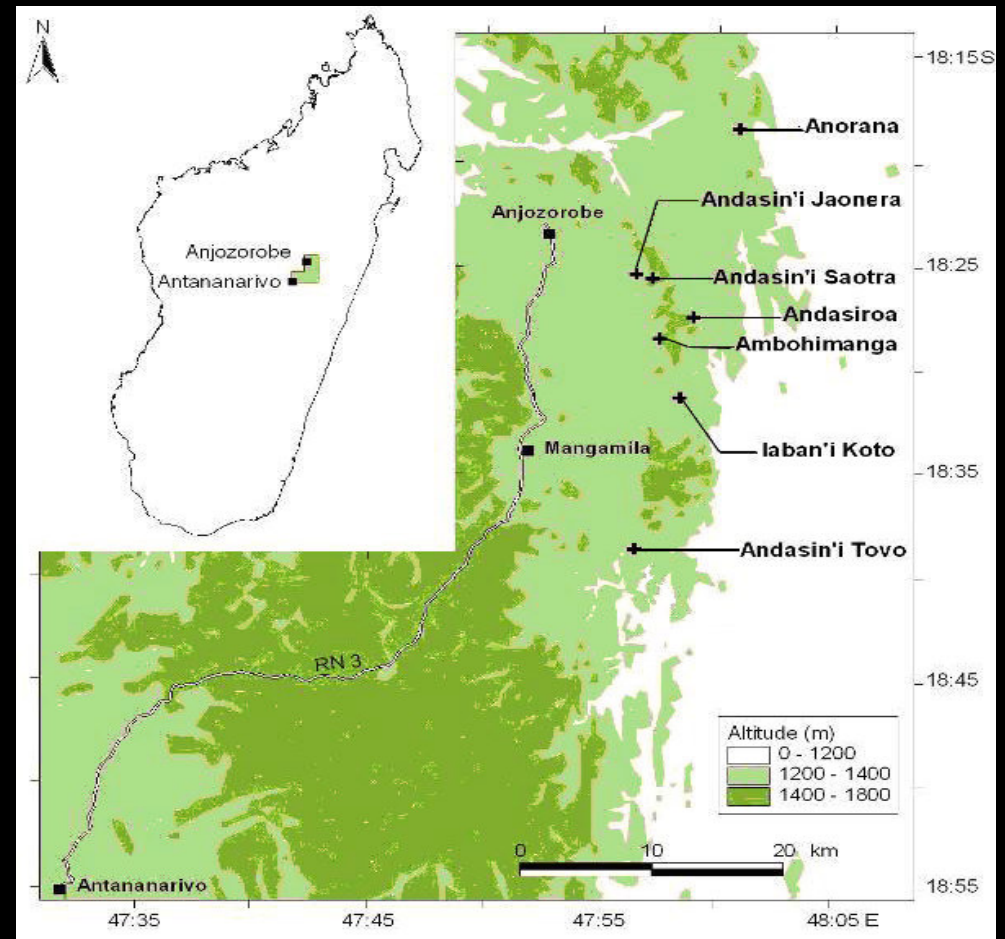
# Research perspectives

- Contact between livestock and wildlife
    - RIFT-OI Madagascar
    - RP-PCP Zimbabwe
  - Experimental infections
    - Viraemia level
    - Viraemia duration
  - Mosquito feeding behaviour
- 



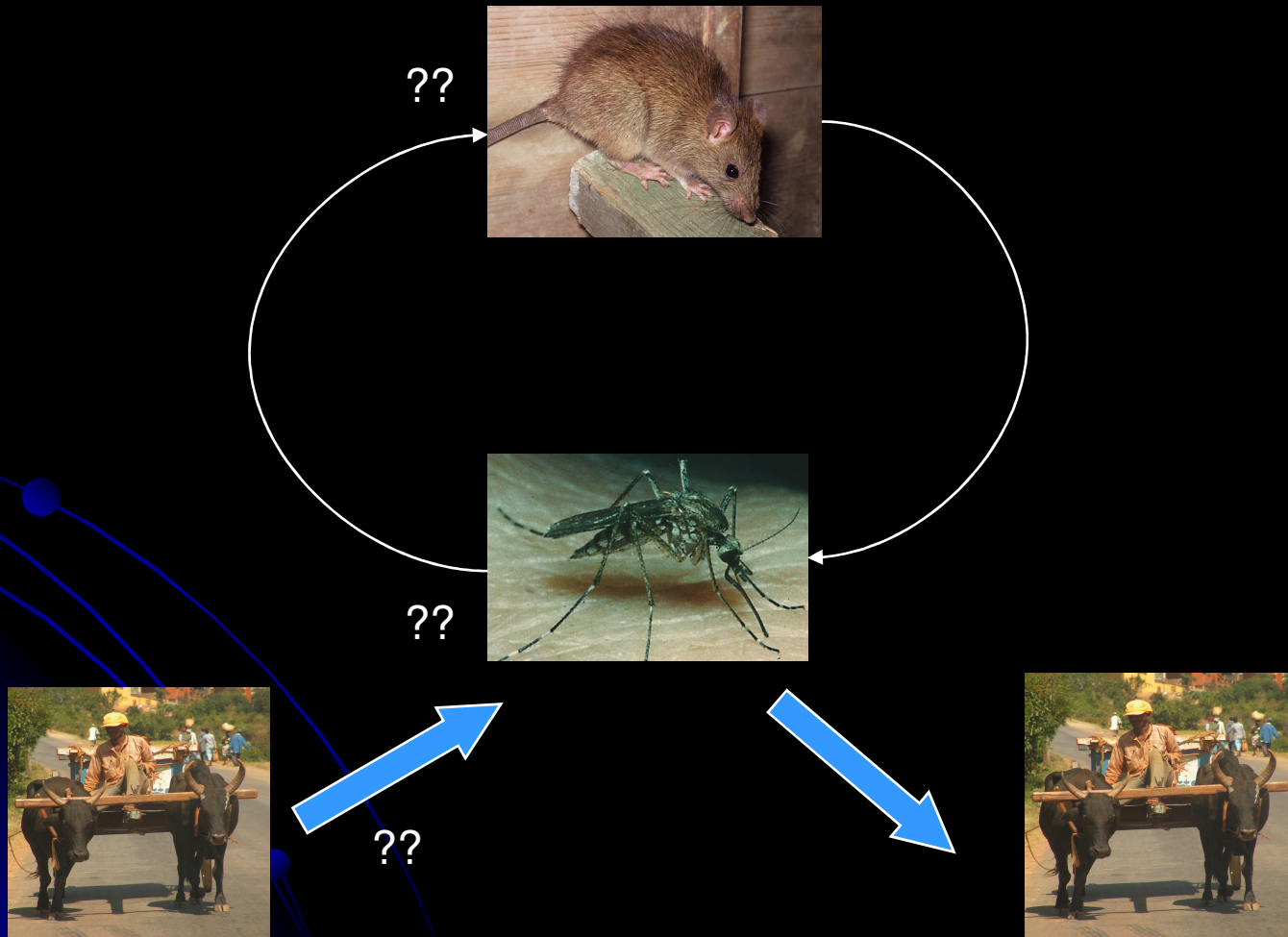
# RIFT-OI project

- Estimation of the incidence on livestock and risk factors
- Identification of potential vectors and population dynamic
- Identification of potential reservoir and population dynamic



# Expected results

- Assumptions on the Anjzorobe ecosystem functioning



# Methodology

Pitfall traps



Sherman traps

# Methodology

1. Standard measures (sex, weight ...)
2. Tissues samples in EDTA,
3. Organs and sera
4. Serological and virological tests



# Overview of ongoing RP-PCP studies in the SEL of Zimbabwe



M. de Garine-Wichatitsky, A. Caron, et al.

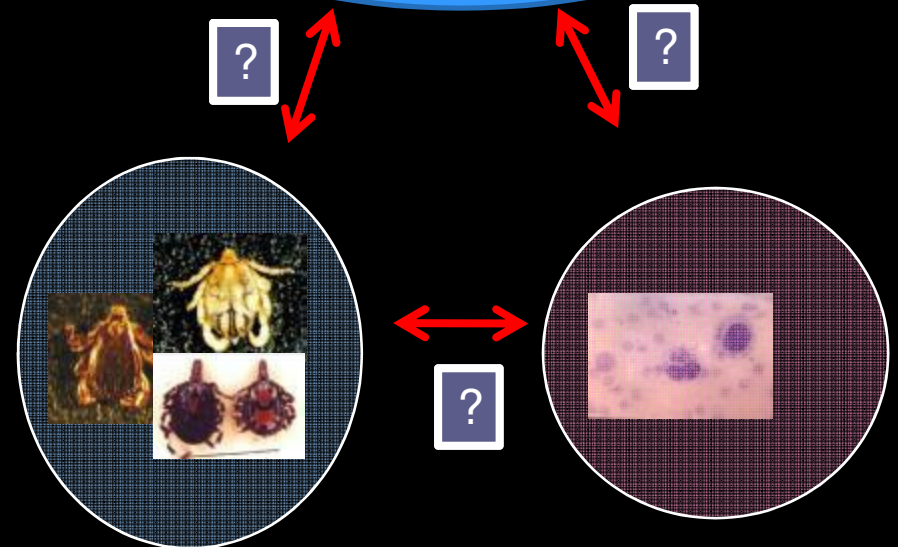
# To understand Host-Pathogen Interactions

- **At the community level:**

- **Multi-hosts**
- **Multi-pathogens**



- **At the wildlife/livestock interface**





## ■ Research activities 2008

- Prevalence of main diseases and parasites in wild ungulates  
40 buffalo, 25 kudu, 40 impala
- Prevalence of main diseases and parasites in domestic ungulates  
120 cattle x 3 seasons 3 sites x 60 cattle/goats/sheep

- Zoonosis:

*Brucellosis*

*BTB*

- Ticks and TBD:

*Theileria spp*

*Boophilus spp/Babesia spp*

*Amblyomma spp/E.ruminantium*

- FMD

- Other viral diseases:

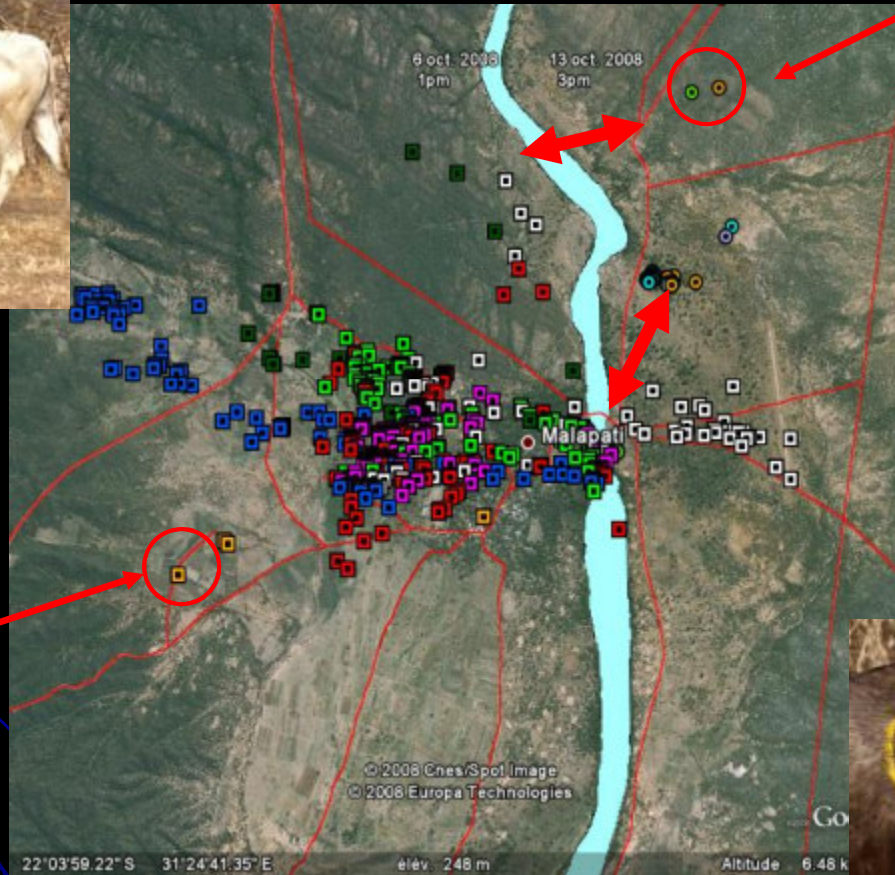
*RVF, LSD, PPR ...*



**EU-PARSEL**

## ■ Research activities in 2008

### • Characterisation of the wildlife/livestock interface



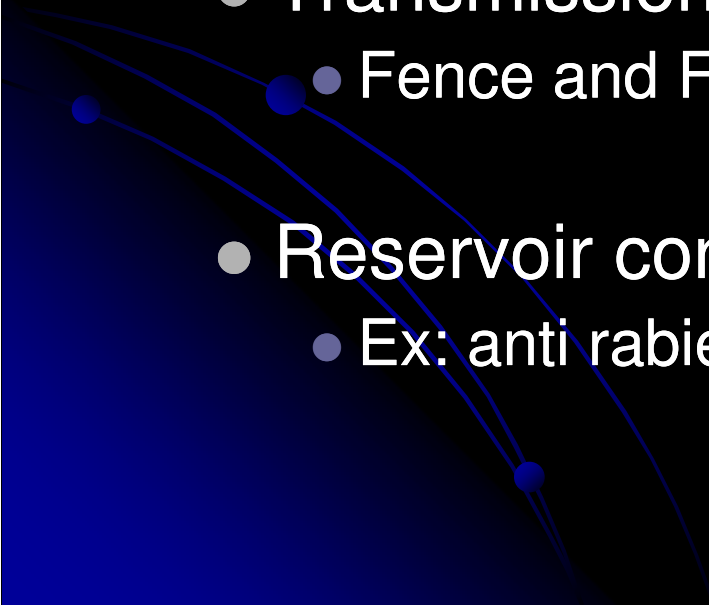
Buffalo

Cattle



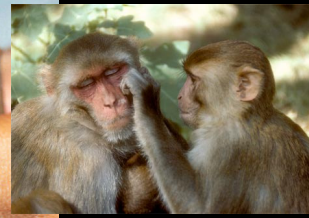
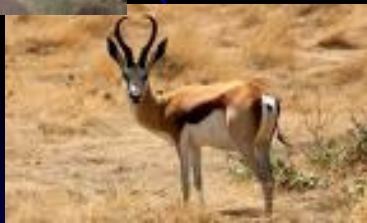
# Consequences in terms of control

Given an identified target-reservoir system

- Target control?
    - Ruminant vaccination
  - Transmission blocking strategy?
    - Fence and FA
  - Reservoir control?
    - Ex: anti rabies fox vaccination
- 

# Conclusion

- More questions than answers !
- Identifying the potential reservoir
  - Field studies
  - Experimental studies
- Understand its role in the epidemiological cycle
  - Modelling
- Adapt the control measures to this cycle



?????

# Contributors

- A. Caron (CIRAD)
- M. de Garine Wichtitsky (CIRAD)
- S. Goodman (Vahatra Association)
- MM. Olive (IPM)
- JM Reynes (IPM)

Thank you for attention !!